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**Broad-sense heritability and genetic gain for powdery mildew resistance in multiple
pseudo-F₂ populations of flowering dogwoods (*Cornus florida* L.)**

Lipi Parikh, M.T. Mmbaga, S. Kodati, M. Blair, D. Hui and G. Meru

10 **Abstract**

11 Powdery mildew caused by *Erysiphe pulchra* is one of the most destructive diseases of flowering dogwoods
12 (*Cornus florida* L.). Control of powdery mildew relies heavily on chemical fungicides. Developing genetic
13 resistance and planting powdery mildew resistant cultivars is a desirable long term control strategy for the disease.
14 Information on inheritance characteristics associated with powdery mildew resistance in *C. florida* is needed to
15 facilitate development of breeding strategies for a new generation of powdery mildew resistant cultivars. The
16 objectives of this study were to determine broad-sense heritability, genetic gain and minimum number of effective
17 genes associated with resistance to powdery mildew in *C. florida*. Progeny segregating for powdery mildew
18 resistance were developed through controlled crosses between susceptible cultivar [Cherokee Princess (CP)],
19 resistant selection (R14) and moderately resistant cultivars [MI9 and Cherokee Brave (CB)]. The parents and
20 progeny were evaluated for resistance/ susceptibility to powdery mildew under greenhouse conditions and estimates
21 for broad-sense heritability, genetic gain and minimum number of effective genes was determined. Broad-sense
22 heritability ranged from 60% to 88% while corresponding narrow-sense heritability ranged from 53% to 86.6%. The
23 highest predicted genetic gain for powdery mildew resistance was obtained in the R14 x CP (78%) and CB x MI9
24 (61%) crosses while the lowest genetic gain was observed in the CP x MI9 (34%) and MI9 x CB (36%) crosses.
25 These results indicate that CB x MI9 and R14 x CP crosses are best parental lines for developing resistance to
26 powdery mildew among the genotypes tested. Use of these parental lines and understanding the gene flow of
27 resistance to powdery mildew in dogwoods will help researchers to develop a targeted breeding approach in
28 generating new generation of resistant cultivars.

29 Keywords: flowering dogwoods, broad-sense heritability, genetic gain, pseudo-F₂, powdery mildew

30 Introduction

31 Flowering dogwoods (*Cornus florida* L.) is native to the U.S and is one of the most popular and economically
32 important ornamental trees in the southeastern U.S with total annual sales of 30 million dollars (United States
33 Department of Agriculture, 2009). The state of Tennessee contributes approximately 23.2% of dogwood sales in the
34 U.S, thus disease constraints that impact dogwood production have ripple effect on the state's economy especially in
35 rural communities. Powdery mildew caused by *Erysiphe pulchra* a (Cook & Peck, Braun & Takamatsu) is the most
36 important constraint in the production of flowering dogwoods (Mmbaga and Sauve, 2004). Powdery mildew fungus
37 produces white mycelia that cover plant foliage thus reducing the total surface available for photosynthesis. This
38 leads to stunted growth, seedling death, reduced aesthetic value and negative impacts on plant sales (Li et al., 2009;
39 Mmbaga, 1998; Mmbaga and Sauve, 2004; Windham et al., 2005). To mitigate these losses, dogwood production
40 requires an effective and consistent strategy for controlling powdery mildew. Currently, disease control relies
41 heavily on the use of chemical fungicides which has in turn increased the production costs and reduced grower
42 profits. As a result of the rising cost in dogwood production, many small-scale nursery growers have reduced or
43 terminated dogwood production (Fulcher and White, 2012; Klingeman et al., 200). In addition, there is a rising
44 concern over the impact of chemical fungicides on the environment. Development and adoption of cultivars resistant
45 to powdery mildew is the most preferred strategy for managing the disease. However, most of the commercially
46 available dogwood cultivars are susceptible to powdery mildew with an exception of a few that are resistant. It is
47 estimated that growers can save approximately \$0.148 per tree and \$593 per acre by planting powdery mildew
48 resistant dogwood cultivars that do not require routine fungicide applications (Gardner et al., 2004; United States
49 Department of Agriculture, 2002). Furthermore, a survey has shown that consumers are willing to pay \$13.35 more
50 for powdery mildew resistant dogwood cultivars than the susceptible cultivars (Gardner et al., 2004). Unfortunately,
51 the few powdery mildew resistant cultivars that are available commercially including 'Cherokee Brave' (Hagan et
52 al., 1998; Mmbaga and Sauve, 2004; Windham et al., 1998), 'Jean's Appalachian Snow', 'Karen's Appalachian
53 Blush', and 'Kay's Appalachian Mist' (Windham et al., 2003) are not resistant across multiple environments and
54 their resistance has not been stable over time (Mmbaga and Sauve, 2004). Therefore, there is an urgent need to
55 develop more cultivars that are resistant to powdery mildew. In addition, there is also a need to understand the total
56 phenotypic variance for the powdery mildew resistance trait as a function of genotype, environment and genotype by
57 environment interactions so as to effectively design targeted breeding programs or selection procedures for genetic

58 improvements (Acquaah, 2011). Heritability is the proportion of the observed phenotypic variation in the progeny
59 that is inherited (Poehlman and Sleper, 1995) while variance due to environmental factors occurs when expression
60 of the genotype is not stable across multiple target environments. In cases where genotype by environment
61 interactions are important, breeding strategies may include the identification of parental lines that can perform well
62 in targeted environments (Fox et al., 1997). Powdery mildew resistance in dogwood has been reported to vary
63 across different environments over multiple years (Mmbaga and Sauve, 2004). Such observations suggest a need for
64 additional studies to better understand the variance components for powdery mildew resistance in dogwood.

65 However, tree crops have long generation time and genetic studies take a long time; furthermore, flowering
66 dogwood have obligatory cross pollination and are highly heterozygous. Fehr (1987) suggested the use of F1 as
67 Pseudo F2 when progeny is derived from controlled crosses for highly heterozygous trees. This was supported by
68 Allard (1999) who explained that when parents are highly heterozygous at many loci due to repeated cross
69 pollination, progeny derived from controlled crosses of such highly heterozygous parents are considered Pseudo F2
70 and are not true F1's. Thus, phenotypic evaluation of F1 as pseudo F2 has been reported in several other cross
71 pollinated crops including mulberry (Banergee et al., 2012), cassava (Aravanopoulous, 2010), tall fescue (Majidi et
72 al., 2009) and willows (Owolade, 2006).

73 Broad sense heritability is the fraction of total phenotypic variance that is genetic due to additive, dominance and
74 epistatic effects (Poehlman and Sleper, 1995). Estimates of broad sense heritability have been useful in
75 understanding the extent to which a trait is influenced by the genotype as opposed to the environment. For example,
76 high broad-sense heritability values found for turf quality, crown density, mowing quality, and genetic color in tall
77 fescue (*Festuca arundinacea* Schreb) (Bokmeyer et al. 2009a; Burton and Devane, 1953) and native prairie
78 junegrass (*Koeleria macrantha* Ledeb) (Clark and Watkins, 2012) indicate that these traits are largely influenced by
79 the genotype as opposed to the environment. Despite the significance of broad sense heritability in trait
80 improvement, such information has not been determined for powdery mildew resistance in dogwoods. Recent
81 reports on narrow sense heritability estimates (70% - 90%) for powdery mildew resistance in flowering dogwoods
82 (Parikh et al., 2015) indicate the importance of additive gene effect (Nyquist, 1991) on this trait.

83 Heritability estimates are useful in estimating genetic gains as prediction parameters of progeny performance
84 (Allard, 1960). While it is easy to breed and predict performance of progeny for qualitatively inherited traits

85 controlled by a few genes (Flor, 1971), predicting progeny performance for quantitatively inherited traits is often
86 difficult and requires estimation of genetic gain (Browning and Frey, 1969; Hooker, 1967). Since powdery mildew
87 resistance in dogwood is quantitatively inherited (Parikh et al., 2015), it is critical to estimate genetic gain for the
88 trait to effectively predict performance of progeny derived from particular crosses.

89 Based on previous crosses between susceptible cultivar [Cherokee Princess (CP)], resistant selection (R14) and
90 moderately resistant cultivars [MI9 and Cherokee Brave (CB)] (Parikh et al., 2015), the objective of the current
91 study was to estimate broad sense heritability, genetic gain and the minimum number of effective genes involved in
92 powdery mildew resistance in dogwood.

93 **Materials and Methods**

94 *Population development.*

95 Controlled crosses were conducted between four parental clones that included powdery mildew resistant (R),
96 moderate resistant (MR) and susceptible (S) genotypes. These crosses were conducted at Tennessee State University
97 Nursery Research Center (TSU-NRC), McMinnville, TN using combinations of R X MR (MI9 x CB), R X S (MI9 x
98 CP and R14 x CP) and their reciprocal crosses (CB x MI9 and CP x MI9) done over a three year period (2004, 2011,
99 and 2012).

100 Controlled crosses were achieved by the hand pollination as described by Reed (1999). After pollination, plant
101 inflorescences remained covered until seeds were harvested. Processing of the harvested seeds involved removal of
102 the red soft seed-coat followed by washing and drying for about 30 days. Seed dormancy was broken by
103 vernalization at 4°C for three months as described by Chouard (1960). Germinants with visible radicals were sown
104 in a Morton's potting mix (Morton's Horticultural Supplies Inc., McMinnville, TN) in 3.75 L pots. MI9 x CB
105 generated 177 progeny plants in 2005, and its reciprocal cross CB x MI9 generated 186 progeny plants in 2013. In,
106 2005 CP x MI9 generated 79 progeny plants whereas MI9 x CP generated 97 progeny plants in 2012. The controlled
107 cross between R14 and CP generated 155 progeny seedlings in 2012. Seedlings of the parental plants and their
108 progeny were maintained in a greenhouse-controlled environment at 26/20°C ±3 (day/night) and 70-90% RH at the
109 TSU-NRC, McMinnville, TN and at the TSU main campus research facilities, Nashville, TN. All plants were
110 fertilized using Nutricote Total™ (Florikan ESA LLC, FL, USA) 18-6-8 (N-P-K) at a rate of 12 g per plant,
111 equivalent to 2.16 g nitrogen, 2.62 g phosphorus and 6.30 g potassium per potted plant and watered daily by
112 overhead sprinkler system to promote powdery mildew disease development.

113 *Plant inoculation.*

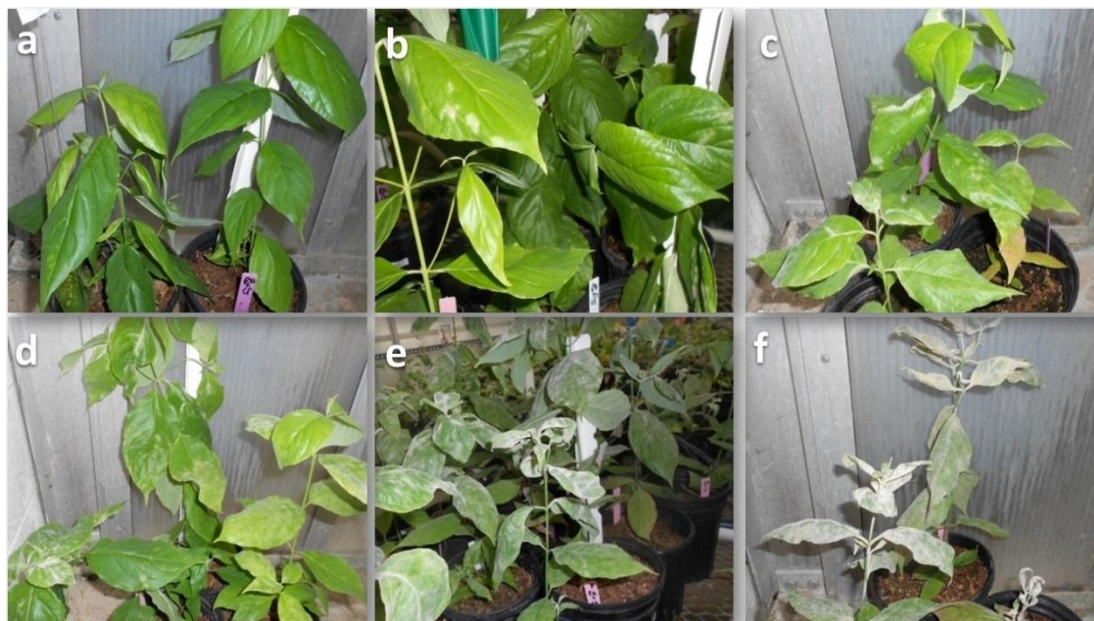
114 Naturally infected plants of susceptible plants were placed in the study area as a source of air-borne inoculum. These
115 plants were placed strategically in the greenhouse to allow even distribution of air-borne spores on the parents and
116 the progeny. Disease severity rating was assessed in 2005, 2012 and 2013 on a scale of 0 to 5 adapted from Horsfall
117 and Barrat (1945), in which a score of 0 represented plants showing 0% symptoms in the foliage, while a score of 1,
118 2, 3, 4 and 5 represented plants showing 1-10%, 11-25%, 26-50%, 51-75% and 76-100% symptoms in the foliage

119 respectively. A visual representation of the disease scale from 0-1 on flowering dogwoods is presented in Figure 1.

120 Disease evaluations were conducted monthly in June, August and September and the highest disease severity was

121 used for subsequent data analysis.

122



123

124 **Figure 1: Powdery mildew disease severity on flowering dogwoods, rating of 0-5 in which 0= No disease**

125 **(a), 1= 1-10% (b), 2= 11-25% (c), 3= 26-50% (d), 4= 51-75% (e), 5= 76-100% (f).**

126

127 *Heritability.*

128 Broad sense heritability was measured using the variance component method (Fehr, 1987). For this method, the

129 formula, $H = \sigma^2_g / \sigma^2_p$; where, σ^2_g = Variance due to genotype (additive, dominance or epistatic) and σ^2_p =

130 Variance due to phenotype was used (Toker, 2004). This analysis used Proc MIXED restricted maximum likelihood

131 (REML estimation of VARCOMP) SAS Institute, NC (2002-2010). Narrow sense heritability was estimated using

132 mid-parent-progeny regression analysis (Poehlman and Sleper, 1995) in which the means of the pseudo F₂ progeny

133 were regressed against mid-parent values from the two parents used in the cross and the slope of the regression line

134 was equal to the narrow sense heritability.

135 *Genetic Gain.*

136 The top 5% of the population was chosen for the selection intensity. Genetic advancement was calculated using the
137 formula from Bonos (2006) and Bonos et al., (2003):

138
$$G = (i) (\sqrt{\sigma_p^2}) (h^2)$$

139 In this formula, i is the selection intensity in top 5% (2.06), σ_p^2 is parental variance among the replicates of parental
140 clone and h^2 is narrow sense heritability calculated as described above.

141 *Minimum number of effective genes.*

142 The minimum number of effective genes controlling powdery mildew resistance in flowering dogwoods was
143 calculated based on the formula by Wright (1968):

144
$$N = (P_1 - P_2)^2 / 8 (\sigma_{F_2}^2 - \sigma_e^2)$$

145 Where, N = Number of genes, P_1 = Mean resistance of parent 1, P_2 = Mean resistance of parent 2, $\sigma_{F_2}^2$ = Variance
146 of F_2 population, σ_e^2 = Environmental variance among the replicates of parental clone pooled for all parental clones.

147 This estimation method makes an assumption that genes have equal effects, no dominance or epistasis present, and
148 that no two loci are on the same chromosome (Poehlman and Sleper, 1995).

149

150

151 **Results and Discussion**

152 *Parental and progeny response to powdery mildew:*

153 There were significant differences in disease severity between the resistant and susceptible parents in all the years
154 (Table 1). For each year, disease severity varied over time (June, August and September). Cherokee Princess
155 showed the highest amount of disease (disease severity ≥ 3) in all the three years indicating highest susceptibility,
156 while R14 selection showed the lowest disease severity (disease severity ≤ 1) indicating highest resistance. As
157 expected, both CB and MI9 selections displayed moderate disease severity (disease severity of 1.5-3.5) indicating
158 moderate resistance. Although CB was released as a highly resistant cultivar (Hagan et al., 1995; Ranney et al.,
159 1994), the results in the current study suggest that this cultivar has moderate resistance as previously reported by
160 Mmbaga and Suave (2004). Based on progeny means for each cross, there was no significant differences in progeny
161 mean disease severity from crosses between R x R, S x R or R x S crosses. The reciprocal crosses did not indicate
162 significant influence of maternal or paternal inheritance on powdery mildew resistance. As reported previously
163 (Parikh et al., 2015), powdery mildew resistance in pseudo F₂ population showed quantitative inheritance with a
164 continuous distribution of disease severity.

165 *Heritability*

166 The estimates for genotype, year and residual effects varied across the 5 crosses (Table 2). Variance due to genotype
167 ranged between 0.56 and 2.06 across all the populations (Table 2). The highest variance due to genotype was from
168 MI9 x CP and R14 x CP followed by MI9 x CB. Variance across the years was generally small and not significant.
169 Similarly, variance resulting from interaction between genotype and year (Genotype*Year) was not significant for
170 CB x MI9, CP x MI9, MI9 x CP and R14 x CP crosses. Variance due to Genotype*Year interaction was highest in
171 MI9 x CB (0.76) and low in the other crosses (Table 2). Residual variance/environmental variance (σ^2_e), ranged
172 from 0.07 to 0.60 and was lower than variance due to genotype (Table 2). Similar results for genotypic variance (0.1
173 - 366.74), variance due to genotype*year interaction (0 - 56) and residual variance (0.07 - 143.08) have been
174 reported for various morphological and fruit color traits in open-pollinated half-sib mango families (Brown et al.,
175 2009).

176 The two-year estimates for broad-sense heritability were 88.9%, 60%, 66%, 80%, and 87.2% for the CB x MI9, MI9
177 x CB, CP x MI9, MI9 x CP and R14 x CP crosses respectively (Table 3). These high broad-sense heritability values
178 (60% - 88.9%) indicate that the total phenotypic variance observed for powdery mildew resistance in flowering
179 dogwoods is mainly accounted for by the genotype (additive, dominance or epistasis effects). Similar high broad-
180 sense heritability has been reported for brown patch resistance in tall fescue (*Festuca arundinacea* Schreb) (74%)
181 (Bokmeyer et al., 2009a). Other quantitative traits with high broad-sense heritability include yield (69% - 99%) in
182 safflower (*Carthamus tinctorius*) (Camas and Esendal, 2006) and leaf size (99.79% - 99.97%) in tobacco (Alekoska
183 and Aleksoki, 2009).

184 Narrow sense heritability (additive gene effects) estimates for powdery mildew resistance in flowering dogwoods
185 using mid-parent progeny means regression ranged from 53.3% to 86.6% (Table 3). The populations that showed
186 high narrow sense heritability of 86.7%, 70.3% and 75% were derived from CB x MI9, MI9 x CP and R14 x CP
187 crosses respectively. High narrow-sense heritability (70% - 97%) has been observed for quantitative traits in tobacco
188 (Alekoska and Alekoski, 2012). Fahlini et al. (2010) also reported estimates of narrow-sense heritability (88% -
189 98%) for agro-morphological traits like grain breadth, plant height, and grain shape and grain length. The level of
190 narrow sense heritability observed for MI9 x CB (53.30%) and CP x MI9 (62.50%) crosses was moderate and is
191 similar to estimates for brown patch resistance (57% - 62%) in tall fescue (Bokmeyer et al., 2009b). These values
192 are also similar to the values (14% - 98%) reported by Stommel and Griesbach (2008) and (76% - 79%) reported by
193 Banerjee et al. (2012) for leaf length and leaf color of capsicum and bacterial leaf spot resistance in mulberry
194 respectively.

195 Estimates for broad-sense heritability in the current study exceeded those of the corresponding narrow-sense
196 heritability. This was expected because broad-sense heritability takes into account total genetic variance (additive +
197 dominance + epistasis effects) as opposed to narrow-sense heritability which only accounts for additive genetic
198 effects (Allard, 1960). The high heritability values reported in this study suggest that most of the phenotypic
199 variance in powdery mildew resistance observed in the five crosses is attributable to genetic effects. Similar results
200 were observed for quantitative traits in tall fescue (Bokmeyer et al., 2009b) and tobacco (Alekoska and Aleksoki,
201 2012), where it was concluded that high broad-sense and narrow sense heritability were indicative of traits being
202 controlled by genetic factors as opposed to environmental effects.

203 *Genetic Gain*

204 Estimates for broad-sense and narrow-sense heritability are useful tools for predicting the genetic gain resulting
205 from a specific bi-parental cross. Estimates for genetic gain calculated using broad-sense heritability take into
206 account all components of genotypic variance (additive, dominance and epistasis effects) and are typically higher
207 than those derived from narrow-sense heritability which accounts only for additive gene effects (Bonos et al., 2003).
208 Since breeders are more interested in the additive gene effects for a given trait, estimation of genetic gain from
209 narrow-sense heritability is prevalent. In our current study, genetic gain for powdery mildew resistance was
210 estimated using values for narrow-sense heritability (Table 4). The genetic gain observed in all the crosses ranged
211 from 33.9% to 77.7%. This genetic gain was calculated with an assumption that the top 5% of the pseudo F₂ plants
212 would be selected for further propagation. The expected genetic gain was highest in the R14 x CP and CB x MI9
213 crosses with an expected gain of 77.7% and 60.9% respectively. High genetic gains of up to 68% have been reported
214 for dollar spot resistance in tall fescue (Bonos et al., 2006). In the current study, low genetic gain was observed for
215 the MI9 x CB (36.1%), CP x MI9 (33.9%) and MI9 x CP (37.3%) crosses. Such low genetic gains are not
216 uncommon in disease resistance traits. For instance, in creeping bent-grass, low genetic gains (28% and 44%) have
217 been previously reported for dollar spot resistance (Bonos, 2006).

218 *Minimum number of effective genes*

219 The minimum number of effective genes contributing to powdery mildew resistance was calculated for all the S x R
220 and R x S crosses and ranged from 1.27 to 3.01 (Table 5). These results support the idea that powdery mildew
221 resistance in flowering dogwood is inherited quantitatively (Parikh et al., 2015). Similar results in number of genes
222 (1.0 - 3.2) has been reported for brown patch resistance in tall fescue (Bokmeyer et al., 2009b). However, the high
223 broad-sense heritability values in this study suggest a possibility of other minor non-additive gene effects. Thus,
224 estimates for the minimum number of effective genes in this current study may be slightly biased. This type of
225 biasness has been reported in turfgrass patho-systems (Bonos et al., 2006; Han et al., 2006).

226 **Conclusion**

227 The findings from this study show high broad-sense heritability for powdery mildew resistance in flowering
228 dogwoods, which strongly suggest that the observed phenotypic variance is largely a function of genetic effects

229 (additive, dominance or epistasis). Conversely, environmental effects are expected to have little influence on
230 expression of this trait. The high narrow-sense heritability for powdery mildew resistance observed in the current
231 study indicates that additive gene effects are more important than non-additive effects (dominance and epistasis) for
232 powdery mildew resistance. The highest genetic gains were observed for the R14 x CP and CB x MI9 crosses which
233 suggest the usefulness of these parental combinations in breeding for powdery mildew resistance in flowering
234 dogwood. The minimum number of effective genes segregating in the S x R and R x S crosses ranged from 1 - 3
235 genes and support previous findings that powdery mildew is inherited quantitatively in flowering dogwoods.

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242 **References**

- 243 Acquaah, G., 2011. *Principles of plant genetics and breeding*. 6. Carlton, Australia: Blackwell publishing.
- 244 Aleksoska, A.K., Aleksoski J., 2012. Heritability of Quantitative Traits in F1 and F2 Progenies of some
245 Domestic and Foreign Tobacco Varieties. *Agriculturae Conspectus Scientificus* 77: 207-210.
- 246 Allard, R.W., 1960. *Principles of plant breeding*. John Wiley & Sons. Inc. New York.
- 247 Allard, R.W., 1999. *Principles of plant breeding*. New York: Wiley.
- 248 Aravanopoulos, F.A., 2010. Contribution of leaf morphometrics in the study of genetic entries in *Salix L.* In (1st
249 Ed) *Electronic Journal of Plant Breeding*, 1320–1328.
- 250 Banerjee, R., Das, N.K., Doss, S.G., Saha, A.K., Bajpai, A.K., Bindroo, B. B., 2012. Narrow sense heritability
251 estimates of bacterial leaf spot resistance in pseudo F2 (F1) population of mulberry (*Morus*
252 spp.). *European Journal of Plant Pathology*. 133: 537 - 544.
- 253 Bokmeyer, J.M., Bonos, S.A., Meyer W.A., 2009a. Broad-sense heritability and stability analysis of brown
254 patch resistance. *HortSci* 44: 289-292.
- 255 Bokmeyer, J.M., Bonos, S.A., Meyer, W.A., 2009b. Inheritance characteristics of brown patch resistance in tall
256 fescue. *CropSci* 49: 2302-2308.
- 257 Bonos, S.A., 2006. Heritability of dollar spot resistance in creeping bentgrass. *Phytopathology* 96: 808-812.
- 258 Bonos, S.A., Clarke, B.B., Meyer, W. A., 2006. Breeding for disease resistance in cool season turfgrass. *Annu.*
259 *Rev. Phytopathol.* 44: 213-234.
- 260 Bonos, S.A., Casler, M.A., Meyer, W.A., 2003. Inheritance of dollar spot resistance in creeping bentgrass.
261 *Crop.Sci.* 43:2189-2196
- 262 Brown, S.J., Schnell, R.J., Ayala-Silva, T., Moore, J.M., Tondo, C.L., Winterstein, M.C., 2009. Broad-sense
263 Heritability Estimates for Fruit Color and Morphological Traits from Open-pollinated Half-sib Mango Families
264 *HortScience* 44: no. 6 1552-1556.
- 265 Browning, J.A., and Frey, K.J., 1969. Multiline cultivars as a means of disease control. *Annu. Rev.*
266 *Phytopathol.* 7355–382.
- 267 Burton, G.W., Devane, E.H., 1953. Estimating heritability in tall fescue (*Festuca arundinacea*) from replicated
268 clonal material.

269 Camas, N., Esendal, E., 2006. Estimates of broad-sense heritability for seed yield and yield components of safflower
270 (Carthamus tinctorius L.). *Hereditas*, 143: 55-7.

271 Chouard, P., 1960. Vernalization and its Relations to Dormancy. *Annual Review of Plant Physiology*, 11, 191-
272 238.

273 Clark, M.D., and Watkins, E., 2012. Broad-sense heritability estimates of turfgrass performance characteristics
274 in native prairie junegrass germplasm. *HortSci* 47: 1228-1233.

275 Fahlini R.A., Khodambashi, M., Houshmand S., Arzani A., 2010. Estimation of the heritability of agro-
276 morphological traits in rice (*Orzya sativa* L.) using F_{2:3} families. *African Journal of Agricultural*
277 *Research* 5: 1297-1303

278 Fehr, W.R., 1987. Theory and technique. *Principles of cultivar development, In Vol 1*. New York: Macmillan
279 Publishing Co.

280 Flor, H.H., (1971). Current status of the gene-for-gene concept. *Annu. Rev. Phthopathol.* 9: 275-296.

281 Fox, P.N., Crossa, J., Ramagosa, I., 1997. Multi-environment testing and genotype x environment interaction.
282 In: Kempton, R.A and Fox, P.N. (eds), *Statistical methods for plant variety evaluation*. Chapman &
283 Hall, p. 117-138

284 Fulcher, A.F., White, S.A., 2012. IPM for select Decidious trees in southeastern US nursery production, Ch.7:
285 *Dogwood-Cornus spp.*

286 Gardner, J.G., Eastwood, D.B., Hall, C.R., Brooker, J.R. 2004. \$0.148/tree. *Production and marketing Reports*,
287 14, 114-119

288 Hagan A.K., Hadin, B., Gilliam, C.H., Keever, G.J., Williams J.D., Eakes, J., 1998. Susceptibility of cultivars
289 of several dogwood taxa to powdery mildew and spot anthracnose. *J. Environ. Hort.* 16: 147-151

290 Hagan, A.K., Gilliam, C., Keever, G., Williams J.D., 1995. Reaction of dogwood selections to powdery mildew.
291 Ornamental Research Report Series No. 10. *Alabama Agricultural Experiment Station*. Auburn
292 University, Auburn, AL. 50 pp

293 Han, Y., Bonos, S.A., Clarke, B.B., Meyer, W.A., 2006. Inheritance of resistance to gray leaf spot disease in
294 perennial ryegrass. *Crop science*, 46, 1143-1148.

295 Hooker, A.L., 1967. The genetics and expression of resistance in plants to rusts of the genus Puccinia. *Annu.*
296 *Rev. Phytopathol.* 5: 163–182.

297Horsfall, J.G., Barrat, R.W., 1945. An improved grading system for measuring plant diseases. (Abstract.)
 298 *Phytopathology* 35:655.

299Klingeman, W.E., Brooker, J.R., Eastwood, D.B., Riley, I.B., Belie, B.K., Knight, P., 2001. Consumer
 300 perceptions of landscape characteristics, disease and pest problems, and the value of powdery mildew
 301 resistant dogwood. *Univ. Tennessee Inst. Agr. Agr. Econ. Res. Serv.* 07-01

302Li, Y., Mmbaga M.T., Windham, A.S., Windham, M.T., Trigliano R.N., 2009. Powdery mildew of dogwoods:
 303 Current status and future prospects. *Plant disease.* 93, 1084-092.

304Majidi, M., Mirlohi, A., & Amini, F., 2009. Genetic variation, heritability and correlations of agro-morphological
 305 traits in tall fescue (*Festuca arundinacea* Schreb.). *Euphytica*, 167(3), 323-331.

306Mmbaga, M.T., 1998. Effect of powdery mildew on growth of dogwood. *Tenn. Nurserymen's Assoc. News.*
 307 8:16–17.

308Mmbaga, M.T., Sauve, R.J., 2004. Multiple disease resistance in dogwoods (*Cornus* spp.) to foliar
 309 pathogens. *Journal of Arboriculture*, 30(2), 101-107

310Nyquist, W., 1991. Estimation of heritability and prediction of selection response in plant populations. *Crit.*
 311 *Rev. Plant Sci.* 10: 235-322.

312Owolade, O.F., 2006. Line x Tester analysis for resistance to cassava anthracnose disease. *World Journal of*
 313 *Agricultural Sciences*, 2, 109–114.

314Parikh, L.P., Mmbaga, M.T., Kodati, S., Zhang G., 2015. Estimation of Narrow sense heritability of powdery
 315 mildew resistance in pseudo-F2 (F1) population of flowering dogwoods (*Cornus florida* L) *European*
 316 *Journal of Plant Path.* DOI 10.1007/s10658-015-0806-5

317Poehlman, J.M., Sleper, D.A., 1995. Breeding field crops. *Iowa State Univ. Press, Ames, IA.*

318Ranney, T.G., Grand, L.F., Knighten, J.L., 1994. Resistance of *Cornus kousa* taxa to dogwood anthracnose and
 319 powdery mildew. *Proc. South. Nurserymen's Assoc. Res. Conf.* 39:212–216.

320Reed, S.M., 1999. Development of a labor-efficient hand pollination procedure for flowering dogwood. *J.*
 321 *Environmental Horticulture* 17, 92-94.

322SAS Institute. 2002-2010. SAS/STAT user's guide, *Version 9.3* Cary, NC.

323Stommel, J.R., Griesbach, R.J., 2008. Inheritance of fruit, foliar, and plant habit attributes in *Capsicum*. *Journal*
 324 *of the American Society for Horticultural Science*, 133, 396-407.

- 325Toker, C., 2004. Estimates of broad-sense heretability for seed yield and yield criteria in faba (*Vicia faba* L.).
326 *Hereditas* 140: 222-225
- 327United States Department of Agriculture. 2009. *2007 census of agriculture*, Washington DC.
- 328United States department of labor. 2002. Producer price index for fungicidal preparations primarily for
329 agriculture, garden or health service. Series PCU2879#7, Bur. Labor Stat., Wash., D.C
- 330Windham, M.T., Witte, W.T., 1998. Naturally occurring resistance to powdery mildew in seedlings of *Cornus*
331 *florida*. *Journal of Environmental Horticulture*, 16, 173-175.
- 332Windham, M.T., Witte, W.T., Trigiano, R.N., 2003. Three white bracted cultivars of *Cornus florida* resistant to powdery
333 mildew. *HortScience* 38:1253–1255
- 334Windham, M.T., Trigiano, R.N., Windham, A.S., 2005. Susceptibility of *Cornus* species to two genera of
335 powdery mildew. *Journal of Environmental Horticulture*, 23(4), 190 - 192.
- 336Wright, S., 1968. *Evolution and the genetics of populations. Vol. 1. Genetic and biométrie foundations. Evolution and*
337 *the genetics of populations. Vol. 1. Genetic and biometrie foundations.*
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- 339

340 **Table 1:** Powdery mildew severity in Cherokee Princess, MI9 and R14 in 2005, 2012 and 2013 in controlled
 341 greenhouse environment

Year of evaluation	Greenhouse progeny evaluation^a	Parents	Mean disease severity^b
2005	GH-1	Cherokee Princess	4.5a
		Cherokee Brave	2.5 cd
		Selection MI9	1.5 ef
2012	GH-1	Cherokee Princess	4.5 a
		Selection R14	1.0 gf
		Selection MI9	1.5 ef
2013	GH-1	Cherokee Princess	3.0 cb
		Selection R14	0.5 g
2013	GH-2	Cherokee Brave	3.5 b
		Selection MI9	2.0 ed

342 LSD 0.05* = 0.661

343 ^a GH-1 – Greenhouse at Tennessee State University Otis Floyd Nursery Research Center,
 344 McMinnville , GH-2- Greenhouse at Tennessee State University main campus (Nashville, TN)

345 ^b Mean disease severity assessed on 0-5 scale in which 0= No disease, 1= 1-10%, 2= 11–25%, 3=
 346 26-50%, 4= 51-75%, 5= 76-100% of the plant covered with powdery mildew. Numbers followed by
 347 same letters are not significantly different (SAS Institute, 2002-2010)

348
 349

350 **Table 2:** Estimates for genotype, year, genotype x year and residual variance components in populations derived
 351 from CB x MI9, MI9 x CB, CP x MI9, MI9 x CP and R14 x CP crosses for the years 2005, 2012 and 2013

Sources of Variation	Variance components ^a				
	CB x MI9	MI9 x CB	CP x MI9	MI9 x CP	R14 x CP
Genotype	0.56	0.75	0.47	1.15	2.06
Year	0	0	0	0.04	0
Genotype*Year	0	0.76	0.15	0.12	0.12
Residual	0.07	0.51	0.12	0.60	0.30

352 ^aVariance components estimated using Proc MIXED REML VARCOMP Method in SAS Inc.

353

354 **Table 3:** Variance components and heritability estimates (broad-sense and narrow-sense) for CB x MI9, MI9 x CB,
 355 CP x MI9, MI9 x CP and R14 x CP crosses

Cross	σ_g^2 ^a	σ_e^2 ^b	σ_p^2 ^c	BSH ^d	NSH ^e
CB x MI9	0.56	0.07	0.63	88.88	86.66
MI9 x CB	0.75	0.51	1.26	60.00	53.30
CP x MI9	1.15	0.60	1.75	66.00	62.50
MI9 x CP	0.47	0.12	0.59	80.00	70.30
R14 x CP	2.06	0.30	2.36	87.20	75.00

356 ^aVariance due to genetic component (σ_g^2)

357 ^bEnvironmental variance (σ_e^2)

358 ^cPhenotypic variance (σ_p^2)

359 ^dBroad-sense heritability estimates ($H = \sigma_g^2 / \sigma_p^2$)

360 ^eNarrow-sense heritability ($h^2 = \text{Slope of mid-parent vs progeny mean regression}$)

361

362 **Table 4:** Predicted gain from selection (Gs) for powdery mildew disease resistance in CB x MI9, MI9 x CB, CP x
 363 MI9, MI9 x CP and R14 x CP crosses. Estimates assumed that only the top 5% of the progeny was be advanced.

Cross	Genetic gain from selection of top 5%	Expected genetic gain in % of the mean	Mean disease severity^a
CB x MI9	1.31	60.9	2.13
MI9 x CB	1.12	36.1	3.10
CP x MI9	1.16	33.9	3.42
MI9 x CP	1.00	37.3	2.68
R14 x CP	2.23	77.7	2.87

364 ^aMean disease severity assessed on 0-5 scale in which 0= No disease, 1= 1-10%, 2= 11-25%, 3= 26-50%, 4= 51-
 365 75%, 5= 76-100% of the plant covered with powdery mildew.

366

367 **Table 5** Estimates for the minimum number of genes controlling powdery mildew resistance in S x R (CP x MI9)
 368 and R x S (MI9 x CP and R14 x CP) crosses.

Cross	Minimum No. of effective Genes^a
CP X MI9	1.27
MI9 X CP	3.01
R14 X CP	2.9

369 ^aMinimum number of effective genes [$N = (P_1 - P_2)^2 / 8 (\sigma_{F2}^2 - \sigma_e^2)$]

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