Tennessee State University

Digital Scholarship @ Tennessee State University

Human Sciences Faculty Research

Department of Human Sciences

7-2019

Synergistic anti-inflammatory effects and mechanisms of combined phytochemicals

Lijuan Zhang Tennessee State University

Carlos Virgous Meharry Medical College

Hongwei Si Tennessee State University

Follow this and additional works at: https://digitalscholarship.tnstate.edu/human-sciences-faculty

Part of the Food Science Commons

Recommended Citation

Lijuan Zhang, Carlos Virgous, Hongwei Si, "Synergistic anti-inflammatory effects and mechanisms of combined phytochemicals", The Journal of Nutritional Biochemistry, Volume 69, 2019, Pages 19-30, ISSN 0955-2863, https://doi.org/10.1016/j.jnutbio.2019.03.009.

This Article is brought to you for free and open access by the Department of Human Sciences at Digital Scholarship @ Tennessee State University. It has been accepted for inclusion in Human Sciences Faculty Research by an authorized administrator of Digital Scholarship @ Tennessee State University. For more information, please contact XGE@Tnstate.edu.

1	Title: Synergistic anti-inflammatory effects and mechanisms of combined phytochemicals
2	Authors: Lijuan Zhang ^a , Carlos Virgous ^b , Hongwei Si ^{a,*}
3	^a Department of Human Sciences, Tennessee State University, Nashville, TN37209, USA
4	^b Animal Care Facility, Meharry Medical College, Nashville, TN 37208, USA
5	
6	* Corresponding author. E-mail: hsi@tnstate.edu. Tel.: +1 615 963 5443
7	
8	
9	
10	
11	
12	Word count for the entire manuscript: 12879
13	The number of figures: 1

14 The number of tables: 1

1 Abbreviations

- 2 AKT, serine/threonine-specific protein kinase
- 3 AMPK, 5'AMP-activated protein kinase
- 4 AP-1, activated activator protein 1 (AP
- 5 CCL2, chemokine (C-C motif) ligand 2
- 6 COX-2, cyclooxygenase-2
- 7 CRP, C-reactive protein
- 8 DHA, docosahexaenoic acid
- 9 EC50, median effective dose
- 10 EGCG, epigallocatechin-3-gallate
- 11 EPA, eicosapentaenoic acid
- 12 H₂O₂, hydrogen peroxide
- 13 HO-1, heme oxygenase-1
- 14 HUVEC, human umbilical vein endothelial cell
- 15 ICAM-1, intercellular adhesion molecule-1
- 16 IFN, interferon
- 17 IL-1, interleukin-1
- 18 iNOS, inducible nitric oxide

- 1 JNKs, c-Jun N-terminal kinases
- 2 LPS, lipopolysaccharides
- 3 MAPK, mitogen-activated protein kinases
- 4 MCP-1, monocyte chemoattractant protein-1 (MCP-1)
- 5 MMPs, matrix metalloproteinases
- 6 NF-kB, nuclear factor kappa-light-chain-enhancer of activated B cells
- 7 NO, nitric oxide
- 8 Nrf2, nuclear factor (erythroid-derived 2)-like 2
- 9 O2•–, superoxide,
- 10 OH•, hydroxyl
- 11 ONOO•–, peroxynitrite
- 12 PEG2, prostaglandin E2
- 13 PUFA, polyunsaturated fatty acid
- 14 SIRT1, sirtuin 1
- 15 ROS, reactive oxygen species
- 16 SOD, superoxide dismutase
- 17 TGF- β , transforming growth factor- β
- 18 TNF- α , tumor necrosis factor- α

1	VCAM-1, vascular cell adhesion molecule-1
2	
3	
4	
5	
6	
7	
8	
9	
10	
11	
12	
13	
14	
15	
16	
17	
18	

1 Abstract

2 The anti-inflammatory effects of phytochemicals, bioactive components from plants having health benefits, have been heavily investigated in the last several decades. However, the gap 3 4 between the high dosage demands (µM) of phytochemicals in vitro studies and the low 5 bioavailability (nM) of most phytochemicals after consuming relevant foods/supplements in 6 humans undermines the application of these phytochemicals in the prevention of chronic 7 inflammation and its related chronic diseases in humans. One of the approaches to bridging this 8 gap is to combine two or more phytochemicals/foods to synergistically prevent chronic 9 inflammation. While increasing combinations of phytochemicals on anti-inflammation studies 10 have been reported, there is no report dedicating why combining two or more phytochemicals 11 synergistically attenuates chronic inflammation. In the present review, we summarized different 12 types of combinations exerting synergistic anti-inflammatory effects such as the combination of 13 phytochemicals from the same foods, and the combination of phytochemicals from different 14 foods/plants. Particularly, we proposed five mechanisms including enhancing the bioavailability of phytochemicals, increasing antioxidant capacity, interacting with gut microbiome and 15 16 targeting same and different signaling pathways, to understand how the combination of 17 phytochemicals exerts synergistic anti-inflammatory effects in cells, animals, and humans. This 18 review provides clues to boost more studies to combine several phytochemicals/foods to reduce 19 chronic inflammation and prevent chronic diseases in humans.

20

21 Key Words: Anti-inflammatory, synergistic, combination, phytochemicals, mechanism

1 **1. Introduction**

2

3 While acute inflammation protects the body by contesting microbial invasion and healing 4 injuries, chronic inflammation attacks critical molecules, cells, and organs to develop various chronic diseases such as cardiovascular disease, diabetes, cancer, and neurological diseases, 5 6 therefore, accelerate aging [1]. Comparing to the signs of acute inflammation such as fever, 7 swelling, redness, and pain, chronic low-grade inflammation is invisible and difficult to notice 8 and is called silent inflammation. The chronic inflammation is characterized by high circulating levels of pro-inflammatory markers including interleukin-1 (IL-1), IL-6, IL-8, IL-13, C-reactive 9 10 protein (CRP), interferon (IFN), transforming growth factor- β (TGF- β), tumor necrosis factor- α 11 (TNF- α) and its soluble receptors and serum amyloid A [2]. These elevated pro-inflammatory 12 molecules further recruit more immune cells such as neutrophils, eosinophils, monocytes, mast 13 cells and platelets to produce more pro-inflammatory molecules as well as nitric oxide (NO), 14 reactive oxygen species (ROS), resulting the damage of structure, function and integrity of lipids, 15 proteins and nucleic acids, and then induce various chronic diseases [3]. The major risk factors 16 of chronic inflammation are aging [1], unhealthy lifestyle involving tobacco use, alcohol use, 17 stress, lack of regular physical activity and obesity [4, 5] as well as environmental pollution [6].

18

19 Strong evidence has been repeatedly presented to support that a healthy lifestyle reduces the 20 risks of chronic inflammation and other chronic diseases [7, 8], particularly selecting healthy 21 foods can significantly prevent chronic diseases from the epidemiological studies. For instance, 22 taking a Mediterranean-like diet was closely associated with relatively lower levels of glucose,

1 lipids, CRP, blood pressure and 10-year cardiovascular risk in men [9]. The Dietary Approaches 2 to Stop Hypertension (DASH) diet, which was originally developed to prevent cardiovascular 3 diseases, significantly reduced circulating CRP and apolipoprotein as well as the rate of 4 cardiovascular disease in humans [10]. Based on these observations, Barry Sears initiated the 5 concept "anti-inflammatory diet" to fight obesity and obesity-induced metabolic syndrome 6 characterized by chronic inflammation about 20 years ago [11]. All these healthy/anti-7 inflammatory diets comprise high consumption of fruits and vegetables (about half of the plate), 8 and these fruits and vegetables contain high levels of phytochemicals, bioactive components 9 from plants having protective effects. These phytochemicals may contribute to the beneficial 10 effects of these healthy diets on the attenuation of chronic inflammation and thereby prevent 11 various chronic diseases [9-11].

12 However, the understanding of the mechanism of these anti-inflammatory diets remains unclear because 1) one food may contain several and even hundreds phytochemicals [12] and our 13 14 diets usually have multiple foods; 2) the complexity of the digestion, absorption, and metabolism 15 of phytochemicals and foods; 3) the disagreement between the high dosage demands of most 16 phytochemicals in in vitro/vivo studies and the low bioavailability of most phytochemicals after 17 consuming relevant foods/supplements in humans [13]. One of the approaches of solving these 18 issues is to combine two or more phytochemicals to investigate if and how the combination 19 synergistically exerts anti-inflammatory effect than the individual chemicals. In the present 20 review, for the first time to our knowledge, we summarized the synergistic anti-inflammatory 21 effects of different types of combinations of phytochemicals, particularly on reducing low-grade 22 chronic inflammation, and proposed possible mechanisms of the synergistic anti-inflammatory 23 effects of the combinations of phytochemicals using cardiovascular disease as a model.

2. Phytochemicals and the major issues of anti-inflammatory research using phytochemicals

3

4 2.1 Phytochemicals

5 Phytochemicals literally mean chemicals from a plant and there is no universal definition 6 acceptable for everyone. However, this term in nutrition is used to describe plant-derived 7 bioactive compounds having the potential health benefits [14, 15]. Most phytochemicals are 8 secondary plant metabolites which are present in a large variety of foods including fruit, 9 vegetables, cereals, nuts, and cocoa/chocolate as well as in beverages including juice, tea, coffee, 10 and wine. More than 1 g of phytochemicals per day is commonly ingested with the diet [16]. 11 According to the chemical structures, phytochemicals can be classified as seven main categories: 12 terpenes, betalains, organosulfides, indoles/glucosinolates/sulfur phenolic compounds, 13 compounds, protein inhibitors, and other organic acids. Table 1 lists some of the phytochemicals 14 now attracting serious scientific attention, identifies food sources and outlines potential anti-15 inflammatory effects.

16

17 Table 1. Classification, food sources, and outlines potential anti-inflammatory effects of

18 phytochemicals

Category			Chemical(s)	Food/Plant resources	Anti-inflammatory Effects
Phenolic compounds	Flavonoids	Flavonols	Fisetin	Strawberries, apples, persimmons, onions, and cucumbers	Inhibits the activity of several pro- inflammatory cytokines, including TNFα, IL- 6, and NF-κB [17]
			Kaempferol	Apples, grapes, tomatoes, green tea, potatoes, onions, broccoli	Reduces the release of TNF-α and IL-1β; Down-regulation the gene and protein expressions of pro- atherogenic molecules, such as E- sel, ICAM-1, VCAM-1 and MCP- 1 [18]

		Colonai		
		Galangin	Alpinia officinarum, Helichrysum aureonitens, and rhizome	Decrease IL-4, IL-5, and IL-13 levels, TNF- α induced p65 nuclear translocation and expression of MCP-1, CXCL10, and VCAM-1 [19]
		Myricetin	Grape, apple, berries, nuts, tea, and red wine	Prevent NF- κ B activation in a monocyte; Inhibits the secretion of IL-6, IL-8 [20]
		Quercetin	Red onions, kale, apples, parsley, sage, tea	Reduce IL-6 and TNF-α levels via modulation of NF-kB [21]
	Flavanones	Hesperetin	lemons and sweet oranges	Reduce inflammatory targets including NF-κB, iNOS, and COX-2, and the markers of chronic inflammation [22]
		Naringenin	Grapefruit, herbs	Decrease the expression and production of TNF-α and MCP-1, suppress NF-κB activation [23]
	Flavones	Apigenin	Parsley, onions, tea, wheat sprouts	Inhibit $TNF\alpha$ -induced $NF \kappa B$ transcriptional activation; inhibits $TNF\alpha$ -induced JNK activation [24]
		Luteolin	Beets, artichokes, leaves, rinds, barks, clover blossom, and ragweed pollen	Active anti-oxidative enzymes, suppress the NF-κB pathway and inhibits pro-inflammatory substances [25]
	Flavan-3-ols	Catechin	Tea, wine, cocoa,	Inhibit TNF- α -induced NF- κ B activity and consequently strongly diminished the secretion of IL-8 [26]
		Epicatechin	Tea, wine, beans, cocoa	Inhibit diet-induced NF-κB activity [27]
		Epigallocatechin gallate	Tea, apple skin, plums, onions, hazelnuts, pecans, and carob powder	Decrease lipid peroxidation, oxidative stress and the production of NO radicals by inhibiting the expression of iNOS; Reduces the activity of NF-κB and AP-1 [28]
		Theaflavin	Black tea	Inhibit TNF-α-mediated activation of IκB kinase and subsequent activation of the IκB-α/NF-κB pathway [29]
		Proanthocyanins	Apples, berries, cocoa-based products, red grapes, red wine	Reduce serum levels of CRP, VCAM-1, and IL-1β [30]
	Anthocyanins	Malvidin	Blueberry, raspberry, black rice, and black soybean	Reduce serum levels of CRP, VCAM-1, and IL-1β [30]
	Isoflavones	Genistein	Lupin, soybeans, kudzu, and psoralea	Reduce peripheral and central NF-kB, NO system and pro-inflammatory cytokine over-activation [31]
		Daidzein	Soybeans	Inhibit the activation of the signal transducer and activator of transcription 1 (STAT-1) [32]
	Chalconoids	Phlorizin	Apple, pear, cherry	Inhibit the levels of NO, PGE2, IL-6, TNF- α , iNOS, and COX-2; suppress the p65 proteins, and decreased phosphorylation in MAPK pathways [33]
Phenolic acids		Ellagic acid	Walnuts, pecans, cranberries, raspberries, strawberries, grapes	Down-regulation of NF–κB, reduce biosynthesis of iNOS and ultimately inhibits the production of NO [34]
		Curcumin	Curcuma longa, Curcuma aromatic, Curcuma zedoaria	suppress the action of IL-6 through the downregulation of STAT3 activation; negatively regulates the action of IL-2;

					suppresses the activation of the transcription factor NF–κB [35]
	Hydroxycinnamic acids		Caffeic acid	Coffee, argan oil, thyme, sage, spearmint	Decrease levels of IL-6, IL-1 β , TNF- α and MCP-1 [36]
	Stilbenoids		Resveratrol	Skin of grapes, blueberries, raspberries, mulberries	Inhibit iNOS and COX-2 via its inhibitory effects on NF- κ B or the activator protein-1 (AP-1) [37]
Terpenes	Carotenoids	Carotenes	Carotene	Sweet potato, carrots, mustard greens, apricots, asparagus, broccoli	Downregulation of iNOS, COX-2, and NADPH oxidase protein and mRNA expression and synergistic inhibition of TNFα secretion [38]
			Lutein	Spinach, kale and yellow carrots	Reduce the level of nuclear NF- κ B, IL-1 β , and Cox-2 [39]
	Monoterpene	Limonene		Oils of citrus, cherries, spearmint	Inhibit the production of ROS; diminished MCP-1 production via NF-kB activation; inhibits cell chemotaxis in a p38 MAPK dependent manner [40]
	Saponins			Vegetables, beans and herbs, soapwort, soaproot, soapbark, and soapberry	Inhibit of COX-2 activity and TNF α production [41]
	Lipids		β-Sitosterol	Vegetable oil, nuts, avocados	Inhibit the phosphorylation of NF- kB [42]
	Triterpenoid		Lupeol	Mango, Acacia visco, abronia villosa	Reduce CD4 + T and CD8 + T cel counts and the level of IL-2, IFN- gamma and IL-4 [43]
Betalains	Betacyanins	Betanin		Beets, Opuntia cactus, Swiss chard	Reduce superoxide anion, TNF- α , and interleukin IL-1 β levels , increase IL-10 levels [43]
	Betaxanthins	Indicaxanthin		Beets, Mirabilis jalapa flowers, prickly pears, red dragonfruit	Inhibit the release of PGE2, NO, IL-1b, and TNF- α , decrease IL-1b TNF- α , iNOS, and COX2 mRNA [44]
Indoles, sulfur compounds	Indole-3-carbinol			Broccoli, cabbage, cauliflower, brussels sprouts, collard greens, and kale	Reduce the production of pro- inflammatory mediators such as NO, IL-6, and IL-1b in through attenuation of the TRIF-dependen signaling pathway; suppress pro- inflammatory cytokine production such as IL-6, TNF- α [45]
	Sulforaphane			Broccoli, Brussels sprouts, and cabbages	Inhibit LPS-stimulated mRNA expression, protein expression, and production of TNF-α, IL-1β, COX-2 and iNOS [46]
	Allicin			Garlic	Increase the phosphorylation of Akt and endothelial nitric oxide synthase (eNOS) [47]
Protein inhibitors	Protease inhibitors			Orange, spinach, rhubarb	Inhibit the downstream portion of the NF-kB pathway; reduce the production of IL-1, TNF- α , IL-6, and IL-10 [48]
Other organic acids	Lactic acid			Koumiss, laban, yogurt, kefir, cottage cheeses	Decrease TNF-α secretion; inhibits NF-kB activation [49]
	Anacardic acid			Cashews, mangoes	Inhibit NF-κB activation; suppress the activation of IκBα kinase; inhibits acetylation and nuclear translocation of p65 [50]

1 2.2 Major issues and solutions of anti-inflammatory research using phytochemicals

2 Although many phytochemicals of each category exert anti-inflammatory effects, the 3 complexity of the digestion, absorption, metabolism, and interactions of phytochemicals and 4 undermines the understanding and application of these anti-inflammatory foods 5 phytochemicals/foods to attenuate chronic inflammation and thereby prevent chronic diseases 6 [51]. For instance, the quantity and composition of phytochemicals in plants are significantly 7 influenced by species, age, part of the plant, cultivation method, harvesting season, preservation 8 method and geographical distribution [52]. Particularly, the range of concentrations of 9 phytochemicals typically used to study mechanisms in cell culture models (1-100 µM, or 10 sometimes higher) is generally higher than the levels in the bloodstream (usually at nM to very 11 low μ M) following consumption of typical doses in foods and supplements [53, 54]. The low 12 bioavailability of phytochemicals may be caused by [13]: 1) special molecular structures such as 13 attached to β -glucosides, high molecular weights, isomeric configuration, hydrophobic and low 14 solubility in the lumen; 2) most phytochemicals do not have the optimal physicochemical 15 properties necessary for passive diffusion, trans-membrane transporters are needed for enhancing 16 their permeability; 3) complex metabolism and interactions between different nutrients and 17 phytochemicals. For instance, after entered into an enterocyte, the phytochemical may be 18 subjected to several phase II enzymes leading to conjugation with methyl (catechol-O-19 methyltransferases-COMT), sulfate (sulphotransferases-SULT) and glucuronyl groups (uridine-20 5'-diphosphate glucuronosyltransferases-UDPGT) and resulted as different chemicals from the 21 original form [55].

However, many phytochemicals and whole foods appear to effectively prevent or ameliorate the symptoms of metabolic syndrome even at low dietary doses in animals [56, 57]

1 and humans [58, 59]. For example, theobromine, a phytochemical from cocoa, synergistically 2 enhanced the anti-hypertensive effect of (-)-epicatechin, the major phytochemicals in cocoa, by 3 increasing the circulating level of (-)-epicatechin in humans [59], indicating whole cocoa 4 (extract) intake is better in reducing blood pressure than that of pure (-)-epicatechin alone. 5 Another study found that co-administrating nutrient mixture (ascorbic acid, selenium, L-lysine, 6 L-proline, L-arginine, N-acetyl cysteine, magnesium, calcium, copper, and manganese) or red 7 onion can increase epigallocatechin-3-gallate (EGCG) level blood by stabilizing EGCG in the 8 lumen in rats and humans [60]. Therefore, whole foods or a combination of several 9 phytochemicals may enhance the health benefits of the phytochemical without increasing 10 dosages. Furthermore, one food may contain multiple even hundreds phytochemicals (for 11 instance, there are about 200 phytochemicals in pomegranate) [12] and one phytochemical may produce many metabolites in the body [51], and these phytochemicals may interact and produce 12 13 more efficient beneficial effects than individual phytochemicals. Indeed, combinations of a 14 couple of phytochemicals synergistically improve osteoporosis [61] and suppress obesity and 15 oxidative stress [62]. In addition, a mixture of wild bilberry, cranberry, elderberry, raspberry, and strawberry exhibited higher antioxidant capacities when compared with the individual berries 16 17 [63]. Breda et al proposed combinations of fruits and vegetables with high levels of 18 phytochemicals to prevent chronic inflammation and chronic diseases [64]. In fact, the major 19 characteristics of famous Mediterranean diet [9] and DASH diet [10] are combinations of high 20 phytochemicals foods such as legumes, olive oil, wine, nuts as well as fruits and vegetables, and 21 the typical Mediterranean diet contains 18 subclasses of phytochemicals and 290 different phytochemicals [65]. Therefore, combine two or more phytochemicals/foods is a reasonable way 22 23 to bridge the gap between the high dosages of demands in cells/animals and the low levels in humans by consuming the foods or supplements, and then reduce chronic inflammation and
prevent chronic diseases in humans.

- 3
- 4

2.3 Measurements of the effects of a combination of two or more phytochemicals

5 Combing two or more phytochemicals is not always enhancing the specific effect. In fact, 6 combination of two or more active chemicals can produce an additive (combined effect is equal 7 to the sum potency of individual components of the mixture), synergistic (combined effect is 8 greater than the sum potency of individual components of the mixture), or antagonistic 9 (combined effect is less than the sum potency of individual components of the mixture) effect 10 [66]. To evaluate and interpret scientifically, in 2006 Chou et al [67] developed the Combination 11 Index (CI): CI \leq 1 indicates synergy; CI =1 means addition; CI > 1 means antagonism. CI is 12 calculated from the below equations.

For binary combination of A and B at 50% activity: $CI_{50} = C_A / IC_{50} (A) + C_B / IC_{50} (B)$, where CI₅₀ is Combination Index for the binary mixture at 50% activity; C_A and C_B is the proportional dose of compound A and compound B (respectively) in the mixture that shows 50% activity; IC₅₀ (A) and IC₅₀ (B) is the single dose of each compound A and B that provides 50% activity.

17 For n-phytochemical combination at x% inhibition:
$${}^{n}(CI)x = \sum_{j=1}^{n} \frac{(D)j}{(Dx)j}$$
, where ${}^{n}(CIx)$ is

a combination index for n compound mixture at x% activity; (D)j is the proportional dose of each
compound in n- phytochemical mixture that shows x% activity; (D_x)j is the single dose of each
compound that provides x% activity.

Based on this CI equation, more and more software (CalcuSyn, Chalice, CompuSyn,
 Combenefit, Genedata Screener, SynergyFinder) have been developed and widely used to
 evaluate the interaction of a combination of chemicals [68].

In this review, CI has been used in numerous articles to screen relevant combinations. CI
and synergy based on the equation have been used, and only the phytochemicals/foods
combinations having synergistic anti-inflammatory effects were discussed.

7

8 **3.** Synergistic anti-inflammation effects of combined phytochemicals

9 3.1 Combination of phytochemicals from the same foods

10 To overcome the controversy that whole food or raw extracts exert beneficial effects but 11 the individual major phytochemicals from the food lose the health benefits, combining several 12 phytochemicals from the same food may exert synergistic anti-inflammatory effects. Indeed, 13 while resveratrol and quercetin can be found from many different foods/plants, both of them are 14 detected from fresh grape skin (50-100 µg/g and 40 µg/g of resveratrol and quercetin 15 respectively) and red wine (7–13 µM and 7.4 µM of resveratrol and quercetin respectively) [69]. A recent study found that combination of resveratrol (120 mg/kg/day) and quercetin (240 16 mg/kg/day) attenuates high fat diet-induced circulating inflammatory markers such as TNF-a, 17 18 IL-6, and monocyte chemoattractant protein-1 (MCP-1) in rats [70]. The same author also 19 reported that combination of resveratrol (2g/kg/day) and quercetin (2g/kg/day) synergistically 20 reversed high fat diet-changed genes of inflammation/immunity compared to the individual 21 chemicals in mice [71]. Similarly, two studies show that combined treatments of flavonoid 22 quercetin and ω-3 polyunsaturated fatty acids (PUFA, available from grape seed) had synergistic anti-inflammatory and antioxidant effects in rats [72, 73]. In a randomized double-masked
controlled human studies, a combination of theobromine and (–)-epicatechin, two phytochemical
from cocoa, synergistically lowered blood pressure and mobilized circulating immune cells [75].
Supplementation with 255 mg/day of a chokeberry flavonoid extract (about 25% anthocyanins,
50% polymeric procyanidins and 9% phenolic acids) for 6 weeks significantly reduced hs-CRP
by 23% in patients after myocardial infarction [74].

7 3.2 Combination of phytochemicals from different foods

8 Most combinations of phytochemicals having synergistic anti-inflammatory effects are 9 combining phytochemicals from different foods/plants. For instance, green tea major 10 phytochemical EGCG (40 nM) and soybean-derived genistein (2 µM) were combined at lower 11 concentrations synergistically inhibits iNOS generation in a murine macrophage RAW264.7 [62]. 12 Rinwa et al reported that co-administration of black pepper-derived piperine (20 mg/kg) with 13 curcumin (100, 200 mg/kg) from spice turmeric synergistically lowered brain TNF- α and caspase 14 3 levels compared to their effects alone in olfactory bulbectomy-induced depression rat [75]. 15 This synergistic anti-inflammatory effects of combined curcumin and piperine have been 16 confirmed in humans [76]. Daily administration of policosanol (majorly from sugar cane) and/or 17 10-dehydrogingerdione at a dose level 10 mg/kg BW synergistically resulted in reducing sP-18 selectin and interferon-gamma (IFN- γ) in dyslipidemic rabbits [77]. Combination of arctigenin 19 and curcumin or curcumin and EGCG synergistically increased bax/bcl-2 ratio and inhibited NF-20 KB levels compared to the individual chemicals, but the most efficient one is the combination of 21 three chemicals together (arctigenin 1 µM, curcumin 5–10 µM, EGCG 40 µM respectively) [78]. Similarly, a combination of curcumin and resveratrol synergistically inhibited inflammation both 22 23 in vitro and in vivo [79, 80]. Ka Lung Cheung et.al reported [81] that curcumin combined with

sulforaphane or phenethyl isothiocyanate synergistically inhibited LPS-induced inflammation in
 RAW 264.7 cells, which was evidenced by the decrease in inducible nitric oxide (iNOS),
 cyclooxygenase-2 (COX-2) protein expression and NO, TNF-α and IL-1 production in the
 medium. Synergistic interaction between Astragali Radix and Rehmanniae Radix in a Chinese
 herbal formula to promote diabetic wound healing [82].

6 Another study found that combination treatment with cocoa polyphenols and ω -3 fatty 7 acids is a promising approach to inhibit inflammation and reduce cardiovascular risk factors 8 associated with aging in humans [83]. Similarly, combinations of eicosapentaenoic acid (EPA) 9 $(0.125 \ \mu\text{M})$ with carnosic acid $(0.2 \ \mu\text{M})$ and lutein $(0.2 \ \mu\text{M})$ caused a synergistic inhibition of 10 prostaglandin E2 (PGE2) release, IL-6 secretion, superoxide and NO production in microglia 11 exposed to lipopolysaccharides (LPS) [84]. 1% curcumin or 0.02% limonin combined with fish 12 oil synergistically suppressed CD4⁺ T-cell proliferation, IL-2 production and NF-KB activity in 13 mice [85]. A combination of lycopene (7.5 μ M), α -tocopherol (1.4 μ M) or ascorbic acid (55 μ M) 14 significantly reduced gene expression and release of the pro-inflammatory cytokines TNF- α and 15 IL-8 but increased anti-inflammatory cytokine IL-10 in human umbilical vein endothelial cell 16 (HUVEC) cells, with similar results of tomato ketchup extracts [86]. A 3-week randomized, 17 double-blind, placebo-controlled, 2×2 factorial clinical study shows that combination of n-3 18 PUFA (1.4 g/d) and plant sterols (2 g/d) per day, synergistically reduced inflammatory markers 19 such as hs-CRP (39%), TNF-α (10%), IL-6 (10.7%) and leukotriene B4 (29.5%) but increased 20 adiponectin by 29.5% [87]. Most importantly, the overall cardiovascular risk was reduced by 21 22.6% (P = 0.006) in the combination group in hyperlipidemic individuals [87]. Lay Saw et.al 22 [88] reported that combinations of curcumin and docosahexaenoic acid (DHA) or EPA have 23 synergistic effects in suppressing LPS-stimulated NO, iNOS, COX-2 in RAW 264.7 cells.

1 We randomly screened combinations (combined two of 20 phytochemicals at various 2 concentrations) to select the combinations having synergistic anti-inflammatory effects using 3 TNF-α-induced monocyte adhesion to endothelial cells. We found that combinations of 4 resveratrol and luteolin (high contents from radicchio, peppers, and celeries) or luteolin and 5 curcumin, at physiological achievable levels, synergistically inhibited TNF-α-induced monocyte 6 adhesion to endothelial cells while the individual chemical did not have significant effects. 7 Moreover, the synergistic anti-inflammatory effects of these two combinations were mediated 8 through regulating intercellular adhesion molecule 1(ICAM-1), MCP-1 and nuclear factor kappa-9 light-chain-enhancer of activated B cells (NF-kB) expressions both in in vitro and in vivo. 10 Particularly, the CI values of these two combinations are around 0.7 in the in vitro studies (data 11 not shown).

12

13 4. Mechanisms of the synergistic anti-inflammation of combined 14 phytochemicals

15 4.1. Enhance the bioavailability/uptake of each other

16

Suganuma et al. found that (–)-epicatechin enhanced the incorporation of EGCG into a human lung cancer cell line PC-9 [89]. Genistein increased cytosolic EGCG by 2- to 5-fold compared with treatment with EGCG only in human colon cancer cells [90]. The same authors also reported that genistein increased the levels of EGCG in the small intestine and plasma following oral dosing of EGCG and genistein in mice [91]. Another study reported that the circulating level of EGCG was significantly increased by co-administrating nutrient mixture

1 because ascorbic acid, selenium, and other nutrients stabilize EGCG in the lumen and help to 2 build up its concentration in the intestine [60]. Similarly, combining DHA (10 µM) with 3 curcumin (10 μ M) significantly enhanced the curcumin uptake in human breast cancer SK-BR-3 4 cells possibly through alteration of membrane lipid composition [92]. This has been observed for 5 DHA in which it enhances the effects of other anti-cancer compounds 5-fluorouracil in colon 6 cancer cells [93], celecoxib in prostate cancer cells [94], and doxorubicin in breast cancer 7 patients [95]. Shoba et al [96] showed that co-administration of piperine and curcumin to humans 8 and rats enhanced the bioavailability of curcumin by 2000% and 154%, respectively. This may 9 be a result of the inhibition of the glucuronidation of curcumin by piperine because curcumin is 10 heavily metabolized in the form of glucuronide conjugates prior to reaching the plasma and 11 piperine is a well-known inhibitor of hepatic and intestinal glucuronidation [97]. The absorption rate of rosmarinic acid in Caco-2 cells is significantly boosted in the presence of luteolin and 12 13 apigenin because luteolin and apigenin inhibit the efflux of rosmarinic acid by inhibiting ABC 14 transporters [98]. Similarly, the bioavailability of quercetin is increased in the presence of 15 proanthocyanidins via forming hydrogen bonds between these chemicals, which possibly contribute to improving the solubility and stability of quercetin [99]. A recent study reported 16 17 that theobromine, a phytochemical from cocoa, synergistically enhanced the anti-hypertensive 18 effect of (–)-epicatechin by increasing the circulating level of (–)-epicatechin in humans [59].

- 19

20 4.2 Synergistically boost antioxidant capacity

The critical roles of oxidative stress on the development of chronic inflammation and chronic diseases have been well recognized [100]. Oxidative stress is a state of imbalance between oxidants and antioxidants in favor of the oxidants, also called the reactive oxygen species (ROS),

1 includes free radicals such as $O_2^{\bullet-}$ (superoxide), ONOO $^{\bullet-}$ (peroxynitrite) and OH $^{\bullet}$ (hydroxyl), 2 and non-radicals such as hydrogen peroxide (H₂O₂). Many individual phytochemicals are found 3 to have the antioxidant capacity of directly scavenging ROS, metal chelating, tempering the 4 mitochondrial respiratory chain, inhibiting certain enzymes as well as increasing endogenous 5 antioxidants enzymes such as superoxide dismutase (SOD), catalase, and glutathione peroxidase 6 both in vitro [101] and in vivo [102]. At the whole food level, Sinha et al [103] tested the total 7 antioxidant capacity (using four assays: ferric reducing antioxidant power, 2,2-diphenyl-1-8 picrylhydrazyl, radical scavenging capacity, and oxygen radical absorbance capacity) of 9 combinations between different foods. Within the same food category, 13% of the tested 10 combinations showed synergistic effect, while across food categories 21% of the tested 11 combinations demonstrated a synergistic effect. The strongest antioxidant capacity is the combination of raspberry and adzuki bean [103]. Fruit with fruit combination also synergistically 12 13 increased antioxidants capacity [104]. For instance, the combination of orange, apple, grape, and 14 blueberry displayed a synergistic effect in antioxidant activity because the median effective dose 15 (EC50) of each fruit after combination was 5 times lower than the EC50 of each fruit alone [105]. 16 Tomatoes contain a matrix of many bioactive components including vitamin C, vitamin E, other 17 carotenoids (a-, beta-, gamma- carotene, lutein), and flavonoids, and mixtures of lycopene and 18 vitamin E appear to have the greatest synergistic antioxidant activity [106].

The possible mechanisms of the synergistic enhancing antioxidant capacity are **1**) protection each other because the combined phytochemicals act in different ways of antioxidation that enable them to protect each other from oxidative agents. For instance, a chain breaking antioxidant scavenges free radicals to protect a singlet oxygen quencher from oxidation and that enables the latter to stay active longer to protect the former against singlet oxygen oxidation

1 [107]; 2) one antioxidant reacts with free radicals or singlet oxygen first to protect the partner 2 from oxidants; 3) one antioxidant is oxidized and becomes a free radical, and this free radical 3 then receive electrons or hydrogen atoms from the other antioxidant to regenerate itself [108]; 4) 4 the chelates metal ions of one antioxidant to allow the oxidant to remain active [109]; 5) the 5 different orientation or position at the water/lipid interface or within the membrane of 6 antioxidants facilitates synergistic interactions [110]; 6) formation of a very strong antioxidant. 7 For example, phytochemical-enhanced heme oxygenase-1 (HO-1) makes stronger antioxidant 8 bilirubin and lead to a higher cellular antioxidant capacity [111]; 7) two or more above six 9 mechanisms may be contributing to the synergistic antioxidants of combined foods or 10 phytochemicals [109, 110, 112].

11 4.3 Target gut microbial profiles and gut integrity

12 The gut microbiome, composed of bacteria, archaea, viruses, and eukaryotic microbes, 13 play critical roles in maintaining healthy physiology and contributing to diseases [113]. Particularly, gut microbiome has critical influences on systemic immune and inflammatory 14 15 components [114, 115]. The interactions between phytochemicals and gut microbiota have been 16 reviewed [116, 117]. A combination of resveratrol (120 mg/kg/day) and quercetin (240 17 mg/kg/day) attenuates high fat diet-induced circulating inflammatory markers such as TNF-a, 18 IL-6 and MCP-1 in rats through regulating gut microbiota, particularly the ratio of 19 Firmicutes/Bacteroidetes ratio and other groups, which are associated with inflammation and 20 immune system [118]. Interestingly, while resveratrol [119] and quercetin alone can regulate the 21 Firmicutes/Bacteroidetes ratio, resveratrol can inhibit the growth of Enterococcus faecalis, and 22 increase the growth of Lactobacillus and Bifidobacterium [119], but quercetin attenuates the 23 growth of Erysipelotrichaceae, Bacillus, Eubacterium cylindroides in rats [120]. In addition, the synergistic anti-inflammatory effect of a combination of resveratrol and quercetin may result from the protective effect of resveratrol on the mucosal barrier integrity [120] which can reduce the circulating endotoxin [121], one of the major stimulators of inflammation. Moreover, a combination of resveratrol and quercetin can up-regulate *Blautia stercoris, Clostridium clariflavum, and Clostridium methylpentosum* [120], which in turns the metabolism of quercetin and other phytochemicals and increased the bioavailabilities of these phytochemicals [117].

7

8 4.4 Target different cells, inflammatory markers, and signaling pathways

9 Inflammation plays a central role in the etiology and development of several vascular 10 diseases including atherosclerosis [122, 123] and ischaemic heart disease [124]. The endothelium 11 is activated by abnormal shear stress, high glucose and result in an increase in permeability to 12 lipoproteins and upregulation of adhesion receptors, and facilitate deposition of lipoproteins in 13 the sub-endothelial space. Activated endothelial cells also promote the recruitment of circulating 14 monocytes by secreting cell-adhesion molecules such as ICAM-1, P-selectin, MCP-1and 15 VCAM-1, these endothelial cells and immune cells further produce chemokines and chemokine 16 receptors such as chemokine (C-C motif) ligand 2 (CCL2), CCL5, CX3C and CCR2, CCR5 to 17 facilitate the transmigration, differentiation and proliferation of monocytes, macrophages, 18 eosinophils and neutrophils as well as T and B lymphocytes. These activated immune cells 19 secrete more pro-inflammatory cytokines such IL-1, IL-4, IL-6, IL-10 and TNF-α, which in turn 20 escalate the inflammation process and generate more ROS as well as NO to damage 21 macromolecules DNA, proteins and oxidize more lipids [125, 126].

22 Resveratrol attenuated monocyte-to-macrophage differentiation, and monocyte infiltration 23 in cells and mice via restoring intracellular glutathione (GSH) levels [127]. Dietary resveratrol

1 significantly lowered levels of mac-3-positive macrophages (a measure of the infiltration of 2 activated macrophages) and reduced ICAM-1, VCAM-1 and MCP-1 expression/levels both in 3 the aorta and plasma as well as reduced the activity of the transcriptional regulator NF-kB in 4 aortic tissues of diabetic mice [128]. However, quercetin can increase the macrophages 5 cholesterol efflux [129] and macrophage migration [130] as well as modulate M1/M2 6 macrophage polarization [131]. These complementary effects of each chemical at least partly 7 contribute to the synergistic anti-inflammatory effects of combined quercetin and resveratrol in 8 animals [70, 71].

9

10 While both curcumin [132] and piperine [133] can regulate common signaling pathways 11 including NF-kB, 5' AMP-activated protein kinase (AMPK), mitogen-activated protein kinases (MAPKs) and nuclear factor (erythroid-derived 2)-like 2(Nrf2)/HO-1, curcumin [134] can 12 13 significantly increase the endothelial nitric oxide level, which is critical for the integrity of 14 endothelial cells and the production of pro-inflammatory markers ICAM-1, VCAM-1 and MCP-1, but there is no report if piperine increases nitric oxide level in endothelial cells. Moreover, the 15 piperine can significantly increase the curcumin bioavailability by inhibiting the hepatic and 16 17 intestinal glucuronidation of curcumin [97]. Moreover, curcumin alone did not have an effect on 18 the IL-10 level, but it significantly increased the piperine-increased IL-10 level in periodontal 19 disease animal model after 15 days treatment [135]. Therefore, these common and different 20 targets complementary contribute to the synergistic anti-inflammation of a combination of 21 piperine and curcumin both in rats [75] and humans [76].

While each phytochemical has its specific interactions on cells, inflammatory markers, and signaling pathways as described above, two or more phytochemicals may also target the same immune and other cells, produce same inflammatory markers by the same pathways. The critical point of the shared targets by different phytochemicals is that a combination of two or more phytochemicals reaches the threshold the level of activating the shared pathway while the individual phytochemical cannot reach this level. This is particularly vital for the low circulating levels of phytochemicals by dietary intake.

8 Because of the key roles in inflammation, macrophages are the major target of anti-9 inflammation research, particularly in vitro studies. Indeed, resveratrol [136], quercetin, 10 kaempferol [137], curcumin [138], genistein [137], luteolin [139], EGCG [28], epicatechin and 11 various food extracts [140, 141] have been shown in inhibiting pro-inflammatory molecules such 12 as TNF-a, IL-1β, IL-6, PGE2 and NO in macrophages. Moreover, resveratrol [142], luteolin 13 [143], and EGCG [144] stimulate anti-inflammatory markers IL-10 and TGF-beta1 expression in 14 macrophages. Quercetin [145] and kaempferol [146] as well as resveratrol [147] inhibits typical 15 pro-inflammatory enzymes, iNOS and COX-2 and therefore decreases NO production via 16 interference with the NF-κB pathway and iNOS/COX-2 induction in Raw264.7 cells. Moreover, 17 luteolin, quercetin, and resveratrol [148, 149] are able to stimulate the expression of the anti-18 inflammatory cytokine IL-10 in macrophages.

Another example is the endothelial cells, the frontline of interaction between environmental factors through circulating blood and inner body, interact with various immune cells to initiate the vascular inflammation. Interestingly, genistein [150, 151], EGCG [152], curcumin [153], resveratrol [154], quercetin [155], luteolin [156] and epicatechin [157] can maintain the integrity of endothelium and reduce adhesion molecules ICAM-1, VCAM-1 and

MCP-1 via regulating endothelial nitric oxide level. These reduced adhesion molecules further
 reduce the adhesion and migrations of monocytes to endothelial cells [132, 158].

3 NF-kB pathway plays a critical role of chronic-inflammation because IL-1 and/or TNF-α-4 activated NF-kB increase production of cytokines, chemokines, and adhesion molecules as well 5 as leukocyte recruitment, and attenuating NF-kB pathway is a key approach to fight chronic 6 inflammation [159]. Indeed, tea extracts and EGCG [160], genistein [161] and resveratrol [162] 7 inhibit inflammation by diminishing NF-kB cascade. Irigenin (3-30µM) inhibits iNOS and 8 COX2 expression via interference with NF- κ B translocation and binding in Raw264.7 cells [163]. 9 Quercetin (~40 µM) has been reported to inhibit IP-10 and MIP-2 expression in intestinal 10 epithelial cells via NF-κB modulation [164]. Curcumin inhibits IL-8, COX-2, and HO-1 11 expression through inhibiting NF-kB in the airway epithelium [165, 166]. Chrysin, ellagic acid, 12 genistein or EGCG at 50 µM reduced IkB-a phosphorylation and diminished IL-8 secretion in 13 intestinal Caco-2 cells [167].

14 The transcription factor nuclear factor (erythroid-derived 2)-like 2(Nrf2), a master 15 regulator of redox homeostasis by regulating the expression and activity of enzymes NADPH, 16 NAD(P)H quinone oxidoreductase 1, glutathione peroxidase, ferritin, heme oxygenase-1 (HO-1), also affects pro-inflammatory mediators including cytokines, COX-2 and iNOS. The most 17 18 investigated phytochemicals including epicatechin [168], resveratrol [169, 170], quercetin [171, 19 172], curcumin [173, 174], EGCG [175], luteolin [176], genistein [177], piperine [178] and 20 apigenin [179] exerts anti-oxidative and anti-inflammatory effects via regulating Nrf2/HO-1 21 pathway [180].

1 Mitogen-activated protein kinases (MAPKs) are a family of serine/threonine protein 2 kinases that mediate fundamental biological processes and cellular responses to external stress 3 signals. Increased activity of MAPK, in particular, p38 MAPK, and their involvement in the 4 regulation of the synthesis of inflammation mediators at the level of transcription and translation 5 make them potential targets for anti-inflammatory therapeutics. The anti-inflammatory activity 6 of curcumin has been associated with a reduction in the activation of p38 MAPK but not c-Jun 7 N-terminal kinases (JNKs) in vivo [181], and resveratrol attenuates inflammation by regulating 8 p38 MAPK [182, 183], therefore, the mutual target p38MAPK mediates the reductions COX-2, 9 IL-6 and IL-8 production by combination of curcumin and resveratrol in normal prostate 10 epithelial [184]. Similarly, the anti-inflammatory actions of quercetin [185], EGCG [186] and 11 curcumin [187] were also associated with activation of the AMPK pathway, suggesting that 12 activation of AMPK may serve as a key mechanism of the phytochemicals anti-inflammatory 13 effects.

14

15 AMPK α 1/sirtuin 1 (SIRT1) signaling pathway exerts its anti-inflammatory effects both in 16 endothelial cells [32], adipocytes [188] and macrophages [189], and AMPK α 1 inhibits the 17 activation of the NF- κ B system [190]. Activation of AMPK α 1 suppresses the synthesis of 18 pro-inflammatory cytokines, such as IL-6 and IL-8 in adipocytes [191]. A combination of 19 resveratrol (120 mg/kg/day) and quercetin (240 mg/kg/day) attenuates high fat diet-induced 20 circulating inflammatory markers such as TNF- α , IL-6, and MCP-1 through regulating 21 AMPK α 1/SIRT1 signaling pathway in rats [70]. This synergistic effect of combined resveratrol 1

2

and quercetin at least results from the shared AMPK α 1/SIRT1 signaling pathway because both resveratrol [192] and quercetin [193] can regulate this pathway alone.

3

4 The complicated interactions between endothelial cells and immune cells and the 5 interactions between these major signaling pathways may contribute to the synergistic anti-6 inflammatory effects of combined phytochemicals. For instance, both resveratrol and quercetin 7 dose-dependently inhibited thrombin-activated endothelial cells, neutrophil migration via 8 regulating MAPK, pMAPK, and JNKs molecules [194], which may partly interpret how the 9 combination of resveratrol and quercetin synergistically reverses high fat diet-induced chronic 10 inflammation in animals [70, 71]. Similarly, a combination of curcumin and sulforaphane 11 synergistically up-regulated HO-1invovling Nrf2, NF-kB and activated activator protein 1 (AP-1) signals and lead to a higher cellular antioxidant capacity by the formation of the very strong 12 13 antioxidant bilirubin [111], and therefore reduced iNOS and COX-2 protein expression and their 14 related inflammatory molecules [81]. Combination of sulforaphane and EGCG synergistically enhanced antioxidant capacity and AP-1, a transcription factor regulates gene expression of 15 inflammation [195]. For instance, both Nrf2/HO-1 pathways and NF-KB pathways mediate the 16 17 anti-inflammatory effect of the combination of curcumin and resveratrol [196].

18

19 5. Conclusions

The synergistic anti-inflammatory effects of combined phytochemicals may be the results of regulating multiple pathways, multiple cells, and inflammatory markers. For the case of combination of curcumin and piperine, in colon and liver, piperine inhibits the hepatic and intestinal glucuronidation of curcumin and increases the curcumin bioavailability, at the same

1 time, the gut microbiome metabolize these chemicals and chemicals, in turn, affect the profiles 2 of the microbiome and finally promote the levels of chemicals in the blood and tissues. After 3 interacting with endothelial cells and various immune cells, the chemicals may directly scavenge 4 the elevated ROS, increase endogenous antioxidants/enzymes such as SOD and/or promotes the 5 Nrf2/HO-1 system to fight oxidative stress. These reduced ROS and/or the chemicals directly 6 further regulates AMPK/SIRT1 or MAPK cascades to attenuate NF-kB pathway. The attenuated 7 NF-kB molecules then deregulate the transcription and translation of pro-inflammatory markers 8 ICAM-1, VCAM-1, MCP-1, TNF-α, IL-1β, IL-6, PGE2 and NO as well as upregulate anti-9 inflammatory molecules such as IL-10 level and endothelial NO, which in turn suppress the 10 proliferation, migration of immune cells and maintain the integrity of endothelial cells to further 11 reduce the production of pro-inflammatory markers and eventually inhibit chronic inflammation (see hypothetical mechanisms using vascular inflammation model as Figure 1). This review 12 13 provides clues to boost more studies to combine several phytochemicals/foods to reduce chronic 14 inflammation and prevent chronic diseases in humans. 15 16 **Disclosure statements** 17 The authors have nothing to disclose. 18 19 Acknowledgments 20 21 This work was supported by grants from the National Institute of Food and Agriculture in the

United States Department of Agriculture (TENX-2011-0255 and TENX-1814-FS to Hongwei Si).

23

1 Figure legends

2

3 Fig. 1. Hypothetical mechanisms of the synergistic anti-inflammatory effect of combined 4 phytochemicals using vascular inflammation as a model. Combination of phytochemicals A 5 and B exerts synergistic anti-inflammatory effects through multiple mechanisms: B increases 6 bioavailability/uptake of A in the blood and tissues by regulating A's metabolism in the gut and 7 liver. At the same time, the interactions between the gut microbiome and phytochemicals result 8 in the changes of the profiles of the microbiome and promotion of the levels of phytochemicals. 9 After interacting with endothelial cells and various immune cells, phytochemicals may directly 10 scavenge the elevated ROS, increase endogenous antioxidants/enzymes such as SOD and/or 11 promotes the Nrf2/HO-1 system to fight oxidative stress. These reduced ROS and/or the chemicals directly further regulates AMPK/SIRT1, Nrf2/HO-1 and/or MAPK cascades to 12 13 attenuate NF-kB pathway in the cytosol. The attenuated NF-kB molecules then transfer into 14 nuclear to regulate the transcription and translation of pro-inflammatory markers ICAM-1, 15 VCAM-1, MCP-1, TNF- α , IL-8, IFN β as well as upregulate anti-inflammatory molecules such as IL-10 level, Nrf2/HO-1, and endothelial NO. These changed molecules, in turn, suppress the 16 17 proliferation, migration of immune cells and maintain the integrity of endothelial cells to further reduce the production of pro-inflammatory markers and eventually inhibit chronic inflammation 18 19 in vasculature.

- 20
- 21

23

22 **References**

Prasad S, Sung B, Aggarwal BB. Age-associated chronic diseases require age-old medicine: role
 of chronic inflammation. Prev Med 2012; 54 Suppl:S29-37.

Ruparelia N, Chai JT, Fisher EA, Choudhury RP. Inflammatory processes in cardiovascular
 disease: a route to targeted therapies. Nature Rev Cardiol 2017; 14.

1	[3]	Ben-Baruch A. Inflammation-associated immune suppression in cancer: The roles played by
2		cytokines, chemokines and additional mediators. Sem Cancer Biol 2006; 16:38-52.
3	[4]	Ford ES, Bergmann MM, Kroger J, Schienkiewitz A, Weikert C, Boeing H. Healthy living is the
4		best revenge findings from the european prospective investigation into cancer and nutrition-
5		potsdam study. Arch Intern Med 2009; 169:1355-62.
6	[5]	Kuroda M, Sakaue H. Adipocyte death and chronic inflammation in obesity. J Med Invest 2017;
7		64:193-6.
8	[6]	Chen J, Zhao Q, Liu BB, Wang J, Xu HB, Zhang Y, Song XM, He B, Huang W. [Airway
9		oxidative stress and inflammation markers in chronic obstructive pulmonary diseases(COPD)
10		patients are linked with exposure to traffic-related air pollution: a panel study]. Zhonghua Yu
11		Fang Yi Xue Za Zhi 2016; 50:411-7.
12	[7]	Arena R, Guazzi M, Lianov L, Whitsel L, Berra K, Lavie CJ, Kaminsky L, Williams M, Hivert
13		MF, Franklin NC et al. Healthy lifestyle interventions to combat noncommunicable disease-a
14		novel nonhierarchical connectivity model for key stakeholders: a policy statement from the
15		american heart association, european society of cardiology, european association for
16		cardiovascular prevention and rehabilitation, and american college of preventive medicine. Mayo
17	101	Clin Proc 2015; 90:1082-103.
18 19	[8]	Avci IA, Nal B, Ayyildiz M. Assessment of chronic disease prevalence, nutritional habits and
20	[0]	healthy lifestyle behaviors in elderly patients. Prog Nutr 2016; 18:26-31.
20 21	[9]	Bonaccio M, Cerletti C, Iacoviello L, de Gaetano G. Mediterranean diet and low-grade subclinical inflammation: the moli-sani study. Endocr Metab Immune 2015; 15:18-24.
$\frac{21}{22}$	[10]	Hodson L, Harnden KE, Roberts R, Dennis AL, Frayn KN. Does the DASH diet lower blood
$\frac{22}{23}$	[10]	pressure by altering peripheral vascular function? J Hum Hypertens 2010; 24:312-9.
23 24	[11]	Sears B: The Zone. New York: Regan books; 1995.
25	[12]	Wu S, Tian L. Diverse phytochemicals and bioactivities in the ancient fruit and modern
26	[12]	functional food pomegranate (Punica granatum). Molecules 2017; 22 (10):E1606.
27	[13]	Rein MJ, Renouf M, Cruz-Hernandez C, Actis-Goretta L, Thakkar SK, da Silva Pinto M.
28	[10]	Bioavailability of bioactive food compounds: a challenging journey to bioefficacy. Br J Clin
29		Pharmacol 2013; 75:588-602.
30	[14]	Jane Higdon, Drake VJ (eds.): An evidence-based approach to phytochemicals and other dietary
31		factors, 2 end; 2013.
32	[15]	Si H, Liu D. Dietary antiaging phytochemicals and mechanisms associated with prolonged
33		survival. J Nutr Biochem 2014; 25:581-91.
34	[16]	Ovaskainen ML, Torronen R, Koponen JM, Sinkko H, Hellstrom J, Reinivuo H, Mattila P.
35		Dietary intake and major food sources of polyphenols in Finnish adults. J Nutr 2008; 138:562-6.
36	[17]	Gupta SC, Tyagi AK, Deshmukh-Taskar P, Hinojosa M, Prasad S, Aggarwal BB.
37		Downregulation of tumor necrosis factor and other proinflammatory biomarkers by polyphenols.
38		Arch Biochem Biophys 2014; 559:91-9.
39	[18]	Rivera Rivera A, Castillo-Pichardo L, Gerena Y, Dharmawardhane S. Anti-breast cancer
40		potential of quercetin via the Akt/AMPK/mammalian target of rapamycin (mTOR) signaling
41	51.03	cascade. PLoS One 2016; 11:e0157251.
42	[19]	Huh JE, Jung IT, Choi J, Baek YH, Lee JD, Park DS, Choi DY. The natural flavonoid galangin
43		inhibits osteoclastic bone destruction and osteoclastogenesis by suppressing NF-kappaB in
44 45		collagen-induced arthritis and bone marrow-derived macrophages. Eur J Pharmacol 2013;
45 46	[20]	698:57-66.
40 47	[20]	Semwal DK, Semwal RB, Combrinck S, Viljoen A. Myricetin: A dietary molecule with diverse biological activities. Nutrients 2016; 8:90.
47 48	[21]	Lesjak M, Beara I, Simin N, Pintać D, Majkić T, Bekvalac K, Orčić D, Mimica-Dukić N.
40 49	[21]	Antioxidant and anti-inflammatory activities of quercetin and its derivatives. J Funct Foods 2018;
5 0		40:68-75.
20		10100 / 21

1 2	[22]	Parhiz H, Roohbakhsh A, Soltani F, Rezaee R, Iranshahi M. Antioxidant and anti-inflammatory properties of the citrus flavonoids hesperidin and hesperetin: an updated review of their molecular
3 4 5	[23]	mechanisms and experimental models. Phytother Res 2015; 29:323-31. Tsai SJ, Huang CS, Mong MC, Kam WY, Huang HY, Yin MC. Anti-inflammatory and antifibrotic effects of naringenin in diabetic mice. J Agric Food Chem 2012; 60:514-21.
6 7	[24]	Funakoshi-Tago M, Nakamura K, Tago K, Mashino T, Kasahara T. Anti-inflammatory activity of structurally related flavonoids, Apigenin, Luteolin and Fisetin. Int Immunopharmacol 2011;
8 9 10	[25]	11:1150-9. Seelinger G, Merfort I, Schempp C. Anti-Oxidant, Anti-inflammatory and anti-allergic activities of luteolin. Planta Medica 2008; 74:1667-77.
10 11 12 13	[26]	Kurbitz C, Heise D, Redmer T, Goumas F, Arlt A, Lemke J, Rimbach G, Kalthoff H, Trauzold A. Epicatechin gallate and catechin gallate are superior to epigallocatechin gallate in growth suppression and anti-inflammatory activities in pancreatic tumor cells. Cancer Sci 2011; 102:728-
14		34.
15 16 17	[27]	Morrison M, van der Heijden R, Heeringa P, Kaijzel E, Verschuren L, Blomhoff R, Kooistra T, Kleemann R. Epicatechin attenuates atherosclerosis and exerts anti-inflammatory effects on diet- induced human-CRP and NFkappaB in vivo. Atherosclerosis 2014; 233:149-56.
18 19	[28]	Zhong Y, Chiou YS, Pan MH, Shahidi F. Anti-inflammatory activity of lipophilic epigallocatechin gallate (EGCG) derivatives in LPS-stimulated murine macrophages. Food Chem
20 21	[29]	2012; 134:742-8. Aneja R, Odoms K, Denenberg AG, Wong HR. Theaflavin, a black tea extract, is a novel anti-
22 23	[30]	inflammatory compound. Critical Care Medicine 2004; 32:2097-103. Zhu Y, Ling W, Guo H, Song F, Ye Q, Zou T, Li D, Zhang Y, Li G, Xiao Y <i>et al</i> . Anti-
23 24 25	[50]	inflammatory effect of purified dietary anthocyanin in adults with hypercholesterolemia: a randomized controlled trial. Nutr Metab Cardiovasc Dis 2013; 23:843-9.
26 27	[31]	Valsecchi AE, Franchi S, Panerai AE, Sacerdote P, Trovato AE, Colleoni M. Genistein, a natural phytoestrogen from soy, relieves neuropathic pain following chronic constriction sciatic nerve
28 29 30 31	[32]	injury in mice: anti-inflammatory and antioxidant activity. J Neurochem 2008; 107:230-40. Yang R, Fang W, Liang J, Lin C, Wu S, Yan S, Hu C, Ke X. Apelin/APJ axis improves angiotensin II-induced endothelial cell senescence through AMPK/SIRT1 signaling pathway. Arch Med Sci 2018; 14:725-34.
32 33 34	[33]	Yuan Y, Shi M, Li L, Liu J, Chen B, Chen Y, An X, Liu S, Luo R, Long D <i>et al.</i> Mesenchymal stem cell-conditioned media ameliorate diabetic endothelial dysfunction by improving mitochondrial bioenergetics via the Sirt1/AMPK/PGC-1alpha pathway. Clin Sci (Lond) 2016;
35 36 37 38	[34]	130:2181-98. Kassim M, Achoui M, Mustafa MR, Mohd MA, Yusoff KM. Ellagic acid, phenolic acids, and flavonoids in Malaysian honey extracts demonstrate in vitro anti-inflammatory activity. Nutr Res 2010; 30:650-9.
39 40 41	[35]	Park YS, Hwang S, Jin YM, Yu Y, Jung SA, Jung SC, Ryu KH, Kim HS, Jo I. CCN1 secreted by tonsil-derived mesenchymal stem cells promotes endothelial cell angiogenesis via integrin alphav beta3 and AMPK. J Cell Physiol 2015; 230:140-9.
42 43	[36]	Chao CY, Mong MC, Chan KC, Yin MC. Anti-glycative and anti-inflammatory effects of caffeic acid and ellagic acid in kidney of diabetic mice. Mol Nutr Food Res 2010; 54:388-95.
44 45	[37]	Gurusamy N, Ray D, Lekli I, Das DK. Red wine antioxidant resveratrol-modified cardiac stem cells regenerate infarcted myocardium. J Cell Mol Med 2010; 14:2235-9.
46 47	[38]	Ciccone MM, Cortese F, Gesualdo M, Carbonara S, Zito A, Ricci G, De Pascalis F, Scicchitano P, Riccioni G. Dietary intake of carotenoids and their antioxidant and anti-inflammatory effects in
48 49 50	[39]	cardiovascular care. Mediators Inflamm 2013; 2013:782137. Li SY, Fung FK, Fu ZJ, Wong D, Chan HH, Lo AC. Anti-inflammatory effects of lutein in retinal ischemic/hypoxic injury: in vivo and in vitro studies. Invest Ophthalmol Vis Sci 2012; 53:5976-
51		84.

1 [40] d'Alessio PA, Ostan R, Bisson JF, Schulzke JD, Ursini MV, Bene MC. Oral administration of d-2 3 limonene controls inflammation in rat colitis and displays anti-inflammatory properties as diet supplementation in humans. Life Sci 2013; 92:1151-6. 4 5 Shao B, Guo H, Cui Y, Ye M, Han J, Guo D. Steroidal saponins from Smilax china and their anti-[41] inflammatory activities. Phytochemistry 2007; 68:623-30. 6 Loizou S, Lekakis I, Chrousos GP, Moutsatsou P. Beta-sitosterol exhibits anti-inflammatory [42] 7 activity in human aortic endothelial cells. Mol Nutr Food Res 2010; 54:551-8. 8 Saleem M. Lupeol, a novel anti-inflammatory and anti-cancer dietary triterpene. Cancer Lett [43] 9 2009; 285:109-15. 10 Allegra M, Ianaro A, Tersigni M, Panza E, Tesoriere L, Livrea MA. Indicaxanthin from cactus [44] pear fruit exerts anti-inflammatory effects in carrageenin-induced rat pleurisy. J Nutr 2014; 11 12 144:185-92. 13 Jiang J, Kang TB, Shim do W, Oh NH, Kim TJ, Lee KH. Indole-3-carbinol inhibits LPS-induced [45] 14 inflammatory response by blocking TRIF-dependent signaling pathway in macrophages. Food 15 Chem Toxicol 2013; 57:256-61. 16 Guo S, Qiu P, Xu G, Wu X, Dong P, Yang G, Zheng J, McClements DJ, Xiao H. Synergistic [46] 17 anti-inflammatory effects of nobiletin and sulforaphane in lipopolysaccharide-stimulated RAW 18 264.7 cells. J Agric Food Chem 2012; 60:2157-64. 19 [47] Chen W, Qi J, Feng F, Wang MD, Bao G, Wang T, Xiang M, Xie WF. Neuroprotective effect of 20 allicin against traumatic brain injury via Akt/endothelial nitric oxide synthase pathway-mediated 21 anti-inflammatory and anti-oxidative activities. Neurochem Int 2014; 68:28-37. 22 Doumas S, Kolokotronis A, Stefanopoulos P. Anti-inflammatory and antimicrobial roles of [48] 23 secretory leukocyte protease inhibitor. Infect Immun 2005; 73:1271-4. 24 Menard S. Lactic acid bacteria secrete metabolites retaining anti-inflammatory properties after [49] 25 intestinal transport. Gut 2004; 53:821-8. 26 [50] Wang B, Mao X. [Protective effect of quercetin against seawater instillation-induced acute lung 27 injury through preventing M1 polarization of macrophages in mice]. Xi Bao Yu Fen Zi Mian Yi 28 Xue Za Zhi 2017; 33:751-5. 29 [51] Borges G, Ottaviani JI, van der Hooft JJJ, Schroeter H, Crozier A. Absorption, metabolism, 30 distribution and excretion of (-)-epicatechin: A review of recent findings. Mol Aspects Med 2018; 31 61:18-30. 32 Lim W, Mudge KW, Vermeylen F. Effects of population, age, and cultivation methods on [52] 33 ginsenoside content of wild American ginseng (Panax quinquefolium). J Agri Food Chem 2005; 34 53:8498-505. 35 Scholl C, Lepper A, Lehr T, Hanke N, Schneider KL, Brockmoller J, Seufferlein T, Stingl JC. [53] Population nutrikinetics of green tea extract. PLoS One 2018; 13:e0193074. 36 37 [54] Clifford MN, van der Hooft JJ, Crozier A. Human studies on the absorption, distribution, 38 metabolism, and excretion of tea polyphenols. Am J Clin Nutr 2013; 98:1619S-30S. 39 Williamson G, Manach C. Bioavailability and bioefficacy of polyphenols in humans. II. Review [55] 40 of 93 intervention studies. Am J Clin Nutr 2005; 81:243S-55S. 41 Babu PV, Si H, Fu Z, Zhen W, Liu D. Genistein prevents hyperglycemia-induced monocyte [56] 42 adhesion to human aortic endothelial cells through preservation of the cAMP signaling pathway 43 and ameliorates vascular inflammation in obese diabetic mice. J Nutr 2012; 142:724-30. 44 [57] Si H, Liu D. Genistein, a soy phytoestrogen, upregulates the expression of human endothelial 45 nitric oxide synthase and lowers blood pressure in spontaneously hypertensive rats. J Nutr 2008; 46 138:297-304. 47 [58] Amiot MJ, Riva C, Vinet A. Effects of dietary polyphenols on metabolic syndrome features in 48 humans: a systematic review. Obes Rev 2016; 17:573-86. 49 [59] Sansone R, Ottaviani JI, Rodriguez-Mateos A, Heinen Y, Noske D, Spencer JP, Crozier A, Merx 50 MW, Kelm M, Schroeter H et al. Methylxanthines enhance the effects of cocoa flavanols on

1		cardiovascular function: randomized, double-masked controlled studies. Am J Clin Nutr 2017;
2	[(0]	105:352-60.
3 4	[60]	Kale A, Gawande S, Kotwal S, Netke S, Roomi W, Ivanov V, Niedzwiecki A, Rath M. Studies on the effects of oral administration of nutrient mixture, quercetin and red onions on the
5		bioavailability of epigallocatechin gallate from green tea extract. Phytother Res 2010; 24 Suppl
6		1:S48-55.
7	[61]	Rayalam S, Della-Fera MA, Baile CA. Synergism between resveratrol and other phytochemicals:
8	[01]	Implications for obesity and osteoporosis. Mol Nutr Food Res 2011; 55:1177-85.
9	[62]	Murakami A, Takahashi D, Koshimizu K, Ohigashi H. Synergistic suppression of superoxide and
10	[02]	nitric oxide generation from inflammatory cells by combined food factors. Mutat Res 2003; 523-
11		524:151-61.
12	[63]	Zafra-Stone S, Yasmin T, Bagchi M, Chatterjee A, Vinson JA, Bagchi D. Berry anthocyanins as
13	[]	novel antioxidants in human health and disease prevention. Mol Nutr Food Res 2007; 51:675-83.
14	[64]	van Breda SGJ, de Kok T. Smart Combinations of Bioactive Compounds in Fruits and
15		Vegetables May Guide New Strategies for Personalized Prevention of Chronic Diseases. Mol
16		Nutr Food Res 2018; 62: doi: 10.1002/mnfr.201700597.
17	[65]	Tresserra-Rimbau A, Medina-Remon A, Perez-Jimenez J, Martinez-Gonzalez MA, Covas MI,
18		Corella D, Salas-Salvado J, Gomez-Gracia E, Lapetra J, Aros F et al. Dietary intake and major
19		food sources of polyphenols in a Spanish population at high cardiovascular risk: The PREDIMED
20		study. Nutr Metab Cardiovas 2013; 23:953-9.
21	[66]	Anh M, Thu Phan, Janet Paterson, Bucknall M, Arcot J. Interactions between phytochemicals
22		from fruits and vegetables: Effects on bioactivities and bioavailability. Cri Rev Food Sci Nutr
23		2018; 58:1310-29.
24	[67]	Chou TC. Theoretical basis, experimental design, and computerized simulation of synergism and
25	1(0)	antagonism in drug combination studies. Pharmacol Rev 2006; 58:621-81.
26	[68]	Bulusu KC, Guha R, Mason DJ, Lewis RP, Muratov E, Kalantar Motamedi Y, Cokol M, Bender
27		A. Modelling of compound combination effects and applications to efficacy and toxicity: state-of-
28 29	[60]	the-art, challenges and perspectives. Drug Discov Today 2016; 21:225-38.
29 30	[69]	Zamin LL, Filippi-Chiela EC, Dillenburg-Pilla P, Horn F, Salbego C, Lenz G. Resveratrol and quercetin cooperate to induce senescence-like growth arrest in C6 rat glioma cells. Cancer Sci
31		2009; 100:1655-62.
32	[70]	Zhao L, Cen F, Tian F, Li MJ, Zhang Q, Shen HY, Shen XC, Zhou MM, Du J. Combination
33	[/0]	treatment with quercetin and resveratrol attenuates high fat diet-induced obesity and associated
34		inflammation in rats via the AMPKalpha1/SIRT1 signaling pathway. Exp Ther Med 2017;
35		14:5942-8.
36	[71]	Zhou M, Wang S, Zhao A, Wang K, Fan Z, Yang H, Liao W, Bao S, Zhao L, Zhang Y <i>et al.</i>
37		Transcriptomic and metabonomic profiling reveal synergistic effects of quercetin and resveratrol
38		supplementation in high fat diet fed mice. J Proteome Res 2012; 11:4961-71.
39	[72]	Camuesco D, Comalada M, Concha A, Nieto A, Sierra S, Xaus J, Zarzuelo A, Galvez J. Intestinal
40		anti-inflammatory activity of combined quercitrin and dietary olive oil supplemented with fish oil,
41		rich in EPA and DHA (n-3) polyunsaturated fatty acids, in rats with DSS-induced colitis. Clin
42		Nutr 2006; 25:466-76.
43	[73]	Denny Joseph KM, Muralidhara. Enhanced neuroprotective effect of fish oil in combination with
44		quercetin against 3-nitropropionic acid induced oxidative stress in rat brain. Prog
45		Neuropsychopharmacol Biol Psychiatry 2013; 40:83-92.
46	[74]	Naruszewicz M, Laniewska I, Millo B, Dluzniewski M. Combination therapy of statin with
47		flavonoids rich extract from chokeberry fruits enhanced reduction in cardiovascular risk markers
48	[<i>7] (</i>]	in patients after myocardial infraction (MI). Atherosclerosis 2007; 194:e179-84.
49 50	[75]	Rinwa P, Kumar A, Garg S. Suppression of neuroinflammatory and apoptotic signaling cascade
50 51		by curcumin alone and in combination with piperine in rat model of olfactory bulbectomy induced depression. PLoS One 2013; 8:e61052.
51		induced depression. I Los One 2015, 0.001052.

- [76] Kaur S, Sharma R, Sarangal V, Kaur N, Prashar P. Evaluation of anti-inflammatory effects of systemically administered curcumin, lycopene and piperine as an adjunct to scaling and root planing: A clinical study. Ayu 2017; 38:117-21.
- [77] Elseweidy MM, Amin RS, Atteia HH, El-Zeiky RR, Al-Gabri NA. New Insight on a
 Combination of Policosanol and 10-Dehydrogingerdione Phytochemicals as Inhibitors for Platelet
 Activation Biomarkers and Atherogenicity Risk in Dyslipidemic Rabbits: Role of CETP and
 PCSK9 Inhibition. Appl Biochem Biotechnol 2018; 186:805-15.
- 8 [78] Wang P, Wang B, Chung S, Wu Y, Henning SM, Vadgama JV. Increased chemopreventive effect
 9 by combining arctigenin, green tea polyphenol and curcumin in prostate and breast cancer cells.
 10 RSC Adv 2014; 4:35242-50.
- [79] Masuelli L, Di Stefano E, Fantini M, Mattera R, Benvenuto M, Marzocchella L, Sacchetti P,
 Focaccetti C, Bernardini R, Tresoldi I *et al.* Resveratrol potentiates the in vitro and in vivo antitumoral effects of curcumin in head and neck carcinomas. Oncotarget 2014; 5:10745-62.
- [80] Majumdar AP, Banerjee S, Nautiyal J, Patel BB, Patel V, Du J, Yu Y, Elliott AA, Levi E, Sarkar
 FH. Curcumin synergizes with resveratrol to inhibit colon cancer. Nutr Cancer 2009; 61:544-53.
- [81] Cheung KL, Khor TO, Kong AN. Synergistic effect of combination of phenethyl isothiocyanate
 and sulforaphane or curcumin and sulforaphane in the inhibition of inflammation. Pharm Res
 2009; 26:224-31.
- [82] Lau KM, Lai KK, Liu CL, Tam JCW, To MH, Kwok HF, Lau CP, Ko CH, Leung PC, Fung KP
 et al. Synergistic interaction between Astragali Radix and Rehmanniae Radix in a Chinese herbal
 formula to promote diabetic wound healing. J Ethnopharmac 2012; 141:250-6.
- [83] Davinelli S, Corbi G, Righetti S, Sears B, Olarte HH, Grassi D, Scapagnini G. Cardioprotection
 by cocoa polyphenols and omega-3 fatty acids: a disease-prevention perspective on aging associated cardiovascular risk. J Med Food 2018; https://doi.org/10.1089/jmf.2018.0002.
- [84] Hadad N, Levy R. Combination of EPA with carotenoids and polyphenol synergistically
 attenuated the transformation of microglia to M1 phenotype via inhibition of NF-kappaB.
 Neuromolecular Med 2017; 19:436-51.
- [85] Kim W, Fan YY, Smith R, Patil B, Jayaprakasha GK, McMurray DN, Chapkin RS. Dietary
 curcumin and limonin suppress CD4+ T-cell proliferation and interleukin-2 production in mice. J
 Nutr 2009; 139:1042-8.
- [86] Hazewindus M, Haenen GR, Weseler AR, Bast A. Protection against chemotaxis in the antiinflammatory effect of bioactives from tomato ketchup. PLoS One 2014; 9:e114387.
- Micallef MA, Garg ML. Anti-inflammatory and cardioprotective effects of n-3 polyunsaturated
 fatty acids and plant sterols in hyperlipidemic individuals. Atherosclerosis 2009; 204:476-82.
- [88] Saw CLL, Huang Y, Kong AN. Synergistic anti-inflammatory effects of low doses of curcumin in combination with polyunsaturated fatty acids: Docosahexaenoic acid or eicosapentaenoic acid. Biochem Pharmac 2010; 79:421-30.
- 38 [89] Suganuma M, Okabe S, Sueoka N, Sueoka E, Matsuyama S, Imai K, Nakachi K, Fujiki H. Green
 39 tea and cancer chemoprevention. Muta Res Fund Mol Mech Mut 1999; 428:339-44.
- 40 [90] Lambert JD, Kwon SJ, Ju JH, Bose M, Lee MJ, Hong JI, Hao XP, Yang CS. Effect of genistein
 41 on the bioavailability and intestinal cancer chemopreventive activity of (-)-epigallocatechin-342 gallate. Carcinogenesis 2008; 29:2019-24.
- 43 [91] Hong J, Lambert JD, Lee SH, Sinko PJ, Yang CS. Involvement of multidrug resistance-associated
 44 proteins in regulating cellular levels of (-)-epigallocatechin-3-gallate and its methyl metabolites.
 45 Biochem Biophysi Res Com 2003; 310:222-7.
- 46 [92] Altenburg JD, Bieberich AA, Terry C, Harvey KA, VanHorn JF, Xu ZD, Davisson VJ, Siddiqui
 47 RA. A synergistic antiproliferation effect of curcumin and docosahexaenoic acid in SK-BR-3
 48 breast cancer cells: unique signaling not explained by the effects of either compound alone. Bmc
 49 Cancer 2011; 11.

- [93] Calviello G, Di Nicuolo F, Serini S, Piccioni E, Boninsegna A, Maggiano N, Ranelletti FO,
 Palozza P. Docosahexaenoic acid enhances the susceptibility of human colorectal cancer cells to
 5-fluorouracil. Cancer Chemothera Pharmac 2005; 55:12-20.
- [94] Narayanan NK, Narayanan BA, Bosland M, Condon MS, Nargi D. Docosahexaenoic acid in
 combination with celecoxib modulates HSP70 and p53 proteins in prostate cancer cells (Retracted article. See vol. 138, pg. 2050, 2016). Int J Cancer 2006; 119:1586-98.
- [95] Bougnoux P, Hajjaji N, Ferrasson MN, Giraudeau B, Couet C, Le Floch O. Improving outcome
 of chemotherapy of metastatic breast cancer by docosahexaenoic acid: a phase II trial. Bri J
 Cancer 2009; 101:1978-85.
- 10[96]Shoba G, Joy D, Joseph T, Majeed M, Rajendran R, Srinivas PS. Influence of piperine on the
pharmacokinetics of curcumin in animals and human volunteers. Planta Med 1998; 64:353-6.
- [97] Reddy L, Odhav B, Bhoola KD. Natural products for cancer prevention: a global perspective.
 Pharmacol Ther 2003; 99:1-13.
- [98] Fale PL, Ascensao L, Serralheiro ML. Effect of luteolin and apigenin on rosmarinic acid
 bioavailability in Caco-2 cell monolayers. Food Funct 2013; 4:426-31.
- [99] Zhao CF, Lei DJ, Song GH, Zhang H, Xu H, Yu LJ. Characterization of water-soluble
 proanthocyanidins of Pyracantha fortuneana fruit and their improvement in cell bioavailable
 antioxidant activity of quercetin. Food Chem 2015; 169:484-91.
- [100] Miller MW, Lin AP, Wolf EJ, Miller DR. Oxidative stress, inflammation, and neuroprogression
 in chronic PTSD. Harv Rev Psychiatry 2018; 26:57-69.
- [101] KL W, RH L. Structure activity relationships of flavonoids in the cellular antioxidant activity
 assay. J Agric Food Chem 2008; 56:8404-11.
- [102] Yahfoufi N, Alsadi N, Jambi M, Matar C. The immunomodulatory and anti-inflammatory role of polyphenols. Nutrients 2018; 10 doi: 10.3390/nu10111618..
- [103] Wang S, Meckling KA, Marcone MF, Kakuda Y, Tsao R. Synergistic, additive, and antagonistic
 effects of food mixtures on total antioxidant capacities. J Agric Food Chem 2011; 59:960-8.
- [104] Huang D, Ou B, Prior RL. The chemistry behind antioxidant capacity assays. J Agric Food Chem
 2005; 53:1841-56.
- [105] Liu RH. Potential synergy of phytochemicals in cancer prevention: mechanism of action. J Nutr
 2004; 134:3479S-85S.
- [106] Shi J, Kakuda Y, Yeung D. Antioxidative properties of lycopene and other carotenoids from 32 tomatoes: synergistic effects. Biofactors 2004; 21:203-10.
- [107] Becker EM, Ntouma G, Skibsted LH. Synergism and antagonism between quercetin and other
 chain-breaking antioxidants in lipid systems of increasing structural organization. Food Chem
 2007; 103:1288-96.
- [108] Vijayalakshmi G, Adinarayana M, Rao PJ. A synergistic approach to kinetic and mechanistic
 studies of regeneration of beta carotene from tert-butoxyl radical induced beta-carotene radical
 cation by chlorogenic acid. Int J Pharm Sci Res 2014; 5:942–50.
- Becker EM, Nissen LR, Skibsted LH. Antioxidant evaluation protocols: Food quality or health
 effects. Eur Food Res Technol 2004; 219:561–71.
- [110] Liang R, Han RM, Fu LM, Ai XC, Zhang JP, Skibsted LH. Baicalin in radical scavenging and its synergistic effect with beta-carotene in antilipoxidation. J Agric Food Chem 2009; 57:7118-24.
- [111] Wang WZW, Smith DLH, Zucker SD. Bilirubin inhibits iNOS expression and NO production in response to endotoxin in rats. Hepatology 2004; 40:424-33.
- Liu Y, Wang Q, Zhang Y, Cui J, Chen G, Xie B, Wu C, Liu H. Synergistic and antagonistic
 effects of salinity and pH on germination in switchgrass (Panicum virgatum L.). PLoS One 2014;
 9:e85282.
- [113] Nagpal R, Mainali R, Ahmadi S, Wang S, Singh R, Kavanagh K, Kitzman DW, Kushugulova A,
 Marotta F, Yadav H. Gut microbiome and aging: Physiological and mechanistic insights. Nutr
 Healthy Aging 2018; 4:267-85.

- [114] Mitchell EL, Davis AT, Brass K, Dendinger M, Barner R, Gharaibeh R, Fodor AA, Kavanagh K.
 Reduced intestinal motility, mucosal barrier function, and inflammation in aged monkeys. J Nutr Health Aging 2017; 21:354-61.
- 4 [115] Takahashi K. Interaction between the intestinal immune system and commensal bacteria and its 5 effect on the regulation of allergic reactions. Biosci Biotech Biochem 2010; 74:691-5.
- [116] Carrera-Quintanar L, Roa RIL, Quintero-Fabian S, Sanchez-Sanchez MA, Vizmanos B, Ortuno Sahagun D. Phytochemicals that influence gut microbiota as prophylactics and for the treatment
 of obesity and inflammatory diseases. Mediators Inflamm 2018: 9734845.
- 9 [117] Selma MV, Espin JC, Tomas-Barberan FA. Interaction between phenolics and gut microbiota:
 10 Role in human health. J Agric Food Chem 2009; 57:6485-501.
- [118] Zhao L, Zhang Q, Ma W, Tian F, Shen H, Zhou M. A combination of quercetin and resveratrol
 reduces obesity in high-fat- diet-fed rats by modulation of gut microbiota. Food Funct 2017;
 8:4644-56.
- [119] Qiao Y, Sun J, Xia S, Tang X, Shi Y, Le G. Effects of resveratrol on gut microbiota and fat
 storage in a mouse model with high-fat-induced obesity. Food Funct 2014; 5:1241-9.
- [120] Eteberria U, Arias N, Boque N, Macarulla MT, Portillo MP, Martinez JA, Milagro FI. Reshaping
 faecal gut microbiota composition by the intake of trans-resveratrol and quercetin in high-fat
 sucrose diet-fed rats. J Nutr Biochem 2015; 26:651-60.
- [121] Terra X, Valls J, Vitrac X, Merrillon JM, Arola L, Ardevol A, Blade C, Fernandez-Larrea J,
 Pujadas G, Salvado J *et al.* Grape-seed procyanidins act as anti-inflammatory agents in
 endotoxin-stimulated RAW 264.7 macrophages by inhibiting NFkB signaling pathway. J Agric
 Food Chem 2007; 55:4357-65.
- [122] Paoletti R, Gotto AM, Jr., Hajjar DP. Inflammation in atherosclerosis and implications for
 therapy. Circulation 2004; 109:III20-6.
- [123] Manduteanu I, Simionescu M. Inflammation in atherosclerosis: a cause or a result of vascular disorders? J Cell Mol Med 2012; 16:1978-90.
- [124] Domingueti CP, Dusse LM, Carvalho M, Gomes KB, Fernandes AP. Hypercoagulability and cardiovascular disease in diabetic nephropathy. Clin Chim Acta 2013; 415:279-85.
- [125] Ruparelia N, Chai JT, Fisher EA, Choudhury RP. Inflammatory processes in cardiovascular disease: a route to targeted therapies. Nat Rev Cardiol 2017; 14:314.
- [126] Robbins CS, Hilgendorf I, Weber GF, Theurl I, Iwamoto Y, Figueiredo JL, Gorbatov R, Sukhova
 GK, Gerhardt LM, Smyth D *et al.* Local proliferation dominates lesional macrophage
 accumulation in atherosclerosis. Nat Med 2013; 19:1166-72.
- [127] Vasamsetti SB, Karnewar S, Gopoju R, Gollavilli PN, Narra SR, Kumar JM, Kotamraju S.
 Resveratrol attenuates monocyte-to-macrophage differentiation and associated inflammation via modulation of intracellular GSH homeostasis: Relevance in atherosclerosis. Free Radic Biol Med 2016; 96:392-405.
- [128] Guo R, Liu B, Wang K, Zhou S, Li W, Xu Y. Resveratrol ameliorates diabetic vascular
 inflammation and macrophage infiltration in db/db mice by inhibiting the NF-kappaB pathway.
 Diab Vasc Dis Res 2014; 11:92-102.
- [129] Sun L, Li E, Wang F, Wang T, Qin Z, Niu S, Qiu C. Quercetin increases macrophage cholesterol
 efflux to inhibit foam cell formation through activating PPARgamma-ABCA1 pathway. Int J Clin
 Exp Pathol 2015; 8:10854-60.
- [130] Cui S, Wu Q, Wang J, Li M, Qian J, Li S. Quercetin inhibits LPS-induced macrophage migration
 by suppressing the iNOS/FAK/paxillin pathway and modulating the cytoskeleton. Cell Adh Migr
 2018; 8:1-12.
- [131] Lu H, Wu L, Liu L, Ruan Q, Zhang X, Hong W, Wu S, Jin G, Bai Y. Quercetin ameliorates
 kidney injury and fibrosis by modulating M1/M2 macrophage polarization. Biochem Pharmacol
 2018; 154:203-12.
- [132] Karimian MS, Pirro M, Johnston TP, Majeed M, Sahebkar A. Curcumin and endothelial function:
 Evidence and mechanisms of protective effects. Curr Pharm Des 2017; 23:2462-73.

1	[133]	Kumar S, Singhal V, Roshan R, Sharma A, Rembhotkar GW, Ghosh B. Piperine inhibits TNF-
2	[155]	alpha induced adhesion of neutrophils to endothelial monolayer through suppression of NF-
$\overline{3}$		kappaB and IkappaB kinase activation. Eur J Pharmacol 2007; 575:177-86.
4	[134]	Santos-Parker JR, Strahler TR, Bassett CJ, Bispham NZ, Chonchol MB, Seals DR. Curcumin
5		supplementation improves vascular endothelial function in healthy middle-aged and older adults
6		by increasing nitric oxide bioavailability and reducing oxidative stress. Aging (Albany NY) 2017;
7		9:187-208.
8	[135]	Guimaraes-Stabili MR, de Aquino SG, de Almeida Curylofo F, Tasso CO, Rocha FRG, de
9		Medeiros MC, de Pizzol JP, Jr., Cerri PS, Romito GA, Rossa C, Jr. Systemic administration of
10		curcumin or piperine enhances the periodontal repair: a preliminary study in rats. Clin Oral
11		Investig 2018: doi: 10.1007/s00784-018-2755-9.
12	[136]	Gonzalez R, Ballester I, Lopez-Posadas R, Suarez MD, Zarzuelo A, Martinez-Augustin O,
13		Sanchez de Medina F. Effects of flavonoids and other polyphenols on inflammation. Crit Rev
14	[107]	Food Sci Nutr 2011; 51:331-62.
15	[137]	Hamalainen M, Nieminen R, Vuorela P, Heinonen M, Moilanen E. Anti-inflammatory effects of
16		flavonoids: genistein, kaempferol, quercetin, and daidzein inhibit STAT-1 and NF-kappaB
17 18		activations, whereas flavone, isorhamnetin, naringenin, and pelargonidin inhibit only NF-kappaB
18 19		activation along with their inhibitory effect on iNOS expression and NO production in activated macrophages. Mediators Inflamm 2007; 2007:45673.
20	[138]	Somchit N, Kimseng R, Dhar R, Hiransai P, Changtam C, Suksamrarn A, Chunglok W,
20	[150]	Chunglok W. Curcumin pyrazole blocks lipopolysaccharide-induced inflammation via
$\frac{21}{22}$		suppression of JNK activation in RAW 264.7 macrophages. Asian Pac J Allergy Immunol 2018;
$\frac{22}{23}$		36:184-90.
24	[139]	Chen D, Bi A, Dong X, Jiang Y, Rui B, Liu J, Yin Z, Luo L. Luteolin exhibits anti-inflammatory
25		effects by blocking the activity of heat shock protein 90 in macrophages. Biochem Biophys Res
26		Commun 2014; 443:326-32.
27	[140]	Bognar E, Sarszegi Z, Szabo A, Debreceni B, Kalman N, Tucsek Z, Sumegi B, Gallyas F, Jr.
28		Antioxidant and anti-inflammatory effects in RAW264.7 macrophages of malvidin, a major red
29		wine polyphenol. PLoS One 2013; 8:e65355.
30	[141]	Park KI, Kang SR, Park HS, Lee DH, Nagappan A, Kim JA, Shin SC, Kim EH, Lee WS, Chung
31		HJ et al. Regulation of proinflammatory mediators via NF-kappaB and p38 MAPK-dependent
32		mechanisms in RAW 264.7 macrophages by polyphenol components isolated from korea lonicera
33	F1 401	japonica THUNB. Evid Based Complement Alternat Med 2012; 2012:828521.
34	[142]	Sharma S, Chopra K, Kulkarni SK, Agrewala JN. Resveratrol and curcumin suppress immune
35 36		response through CD28/CTLA-4 and CD80 co-stimulatory pathway. Clin Exp Immunol 2007;
30 37	[143]	147:155-63. Zhang BC, Li Z, Xu W, Xiang CH, Ma YF. Luteolin alleviates NLRP3 inflammasome activation
38	[143]	and directs macrophage polarization in lipopolysaccharide-stimulated RAW264.7 cells. Am J
39		Transl Res 2018; 10:265-73.
40	[144]	Xu Z, Wei C, Zhang RU, Yao J, Zhang D, Wang L. Epigallocatechin-3-gallate-induced inhibition
41	[1.1]	of interleukin-6 release and adjustment of the regulatory T/T helper 17 cell balance in the
42		treatment of colitis in mice. Exp Ther Med 2015; 10:2231-8.
43	[145]	Lee S, Park HS, Notsu Y, Ban HS, Kim YP, Ishihara K, Hirasawa N, Jung SH, Lee YS, Lim SS
44		et al. Effects of hyperin, isoquercitrin and quercetin on lipopolysaccharide-induced nitrite
45		production in rat peritoneal macrophages. Phytother Res 2008; 22:1552-6.
46	[146]	Wang GF, Wu ZF, Wan L, Wang QT, Chen FM. Influence of baicalin on the expression of
47		receptor activator of nuclear factor-kappaB ligand in cultured human periodontal ligament cells.
48		Pharmacology 2006; 77:71-7.
49	[147]	Yu W, Tao M, Zhao Y, Hu X, Wang M. 4'-Methoxyresveratrol alleviated AGE-induced
50		inflammation via RAGE-mediated NF-kappaB and NLRP3 inflammasome pathway. Molecules
51		2018; 23;1447.

1 2	[148]	Comalada M, Ballester I, Bailon E, Sierra S, Xaus J, Galvez J, de Medina FS, Zarzuelo A. Inhibition of pro-inflammatory markers in primary bone marrow-derived mouse macrophages by
$\frac{2}{3}$		naturally occurring flavonoids: analysis of the structure-activity relationship. Biochem Pharmacol
4		2006; 72:1010-21.
5	[149]	Imler TJ, Jr., Petro TM. Decreased severity of experimental autoimmune encephalomyelitis
6		during resveratrol administration is associated with increased IL-17+IL-10+ T cells, CD4(-) IFN-
7		gamma+ cells, and decreased macrophage IL-6 expression. Int Immunopharmacol 2009; 9:134-
8 9	[150]	43. Si H, Liu D. Isoflavone genistein protects human vascular endothelial cells against tumor necrosis
10	[150]	factor-alpha-induced apoptosis through the p38beta mitogen-activated protein kinase. Apoptosis
11		2009; 14:66-76.
12	[151]	Si H, Yu J, Jiang H, Lum H, Liu D. Phytoestrogen genistein up-regulates endothelial nitric oxide
13	L - J	synthase expression via activation of cAMP response element-binding protein in human aortic
14		endothelial cells. Endocrinology 2012; 153:3190-8.
15	[152]	Liu S, Sun Z, Chu P, Li H, Ahsan A, Zhou Z, Zhang Z, Sun B, Wu J, Xi Y et al. EGCG protects
16		against homocysteine-induced human umbilical vein endothelial cells apoptosis by modulating
17		mitochondrial-dependent apoptotic signaling and PI3K/Akt/eNOS signaling pathways. Apoptosis
18	54 503	2017; 22:672-80.
19	[153]	Zhang Z, Li K. Curcumin attenuates high glucose-induced inflammatory injury through the
20 21		reactive oxygen species-phosphoinositide 3-kinase/protein kinase B-nuclear factor-kappaB signaling pathway in rat thoracic aorta endothelial cells. J Diabetes Investig 2018; 9:731-40.
22	[154]	Arunachalam G, Yao H, Sundar IK, Caito S, Rahman I. SIRT1 regulates oxidant- and cigarette
23	[134]	smoke-induced eNOS acetylation in endothelial cells: Role of resveratrol. Biochem Biophys Res
24		Commun 2010; 393:66-72.
25	[155]	Tribolo S, Lodi F, Winterbone MS, Saha S, Needs PW, Hughes DA, Kroon PA. Human
26		metabolic transformation of quercetin blocks its capacity to decrease endothelial nitric oxide
27		synthase (eNOS) expression and endothelin-1 secretion by human endothelial cells. J Agric Food
28		Chem 2013; 61:8589-96.
29	[156]	Jia Z, Nallasamy P, Liu D, Shah H, Li JZ, Chitrakar R, Si H, McCormick J, Zhu H, Zhen W et al.
30		Luteolin protects against vascular inflammation in mice and TNF-alpha-induced monocyte
31 32		adhesion to endothelial cells via suppressing IKappaBalpha/NF-kappaB signaling pathway. J Nutr Biochem 2015; 26:293-302.
33	[157]	Ramirez-Sanchez I, Maya L, Ceballos G, Villarreal F. (-)-Epicatechin induces calcium and
34	[137]	translocation independent eNOS activation in arterial endothelial cells. Am J Physiol Cell Physiol
35		2011; 300:C880-7.
36	[158]	Pendurthi UR, Rao LV. Resveratrol suppresses agonist-induced monocyte adhesion to cultured
37		human endothelial cells. Thromb Res 2002; 106:243-8.
38	[159]	Lawrence T. The nuclear factor NF-kappaB pathway in inflammation. Cold Spring Harb Perspect
39		Biol 2009; 1:a001651.
40	[160]	Yang FJ, Oz H, Barve S, Devilliers WJ, McClain CJ, Varilek G. The green tea polyphenol, (-)-
41		epigallocatechin-3-gallate blocks nuclear factor-kappa B activation by inhibiting IkappaB kinase
42 43	[161]	activity in the intestinal epithelial cell line, IEC-6. Gastroenterology 2001; 120:A188-A.
43 44	[161]	Li JC, Li J, Yue Y, Hu YP, Cheng WX, Liu RX, Pan XH, Zhang P. Genistein suppresses tumor necrosis factor alpha-induced inflammation via modulating reactive oxygen species/Akt/nuclear
45		factor kappa B and adenosine monophosphate-activated protein kinase signal pathways in human
46		synoviocyte MH7A cells. Drug Des Dev Ther 2014; 8:315-23.
47	[162]	Wu M, Gu JH, Mei SQ, Xu DC, Jing Y, Yao Q, Chen MH, Yang M, Chen SX, Yang B <i>et al.</i>
48		Resveratrol delays polycystic kidney disease progression through attenuation of nuclear factor
49		kappa B-induced inflammation. Nephrol Dial Transpl 2016; 31:1826-34.

- [163] Ahn KS, Noh EJ, Cha KH, Kim YS, Lim SS, Shin KH, Jung SH. Inhibitory effects of Irigenin
 from the rhizomes of Belamcanda chinensis on nitric oxide and prostaglandin E(2) production in
 murine macrophage RAW 264.7 cells. Life Sci 2006; 78:2336-42.
- [164] Ruiz PA, Braune A, Izlwimmer GH, Quintanilla-Fend L, Haller D. Quercetin inhibits TNF-α
 induced NF-kappaB transcription factor recruitment to proinflammatory gene promoters in
 murine intestinal epithelial cells. J Nutr 2007; 137:1208-15.
- [165] Shishodia S, Potdar P, Gairola CG, Aggarwal BB. Curcumin (diferuloylmethane) down-regulates
 cigarette smoke-induced NF-kappaB activation through inhibition of IkappaBalpha kinase in
 human lung epithelial cells: correlation with suppression of COX-2, MMP-9 and cyclin D1.
 Carcinogenesis 2003; 24:1269-79.
- [166] SK B, D M, LA J, IL M, I R. Curcumin induces glutathione biosynthesis and inhibits NF-kappaB
 activation and interleukin-8 release in alveolar epithelial cells: mechanism of free radical
 scavenging activity. Antioxid Redox Signal 2005; 7:32-41.
- [167] Romier B, Van De Walle J, During A, Larondelle Y, Schneider YJ. Modulation of signalling
 nuclear factor-kappaB activation pathway by polyphenols in human intestinal Caco-2 cells. Br J
 Nutr 2008; 100:542-51.
- [168] Ahmed SM, Luo L, Namani A, Wang XJ, Tang X. Nrf2 signaling pathway: Pivotal roles in inflammation. Biochim Biophys Acta Mol Basis Dis 2017; 1863:585-97.
- [169] Bigagli E, Cinci L, Paccosi S, Parenti A, D'Ambrosio M, Luceri C. Nutritionally relevant
 concentrations of resveratrol and hydroxytyrosol mitigate oxidative burst of human granulocytes
 and monocytes and the production of pro-inflammatory mediators in LPS-stimulated RAW 264.7
 macrophages. Int Immunopharmacol 2017; 43:147-55.
- [170] Kumar A, Singh CK, Lavoie HA, Dipette DJ, Singh US. Resveratrol restores Nrf2 level and
 prevents ethanol-induced toxic effects in the cerebellum of a rodent model of fetal alcohol
 spectrum disorders. Mol Pharmacol 2011; 80:446-57.
- [171] Li X, Wang H, Gao Y, Li L, Tang C, Wen G, Zhou Y, Zhou M, Mao L, Fan Y. Protective effects
 of quercetin on mitochondrial biogenesis in experimental traumatic brain injury via the Nrf2
 signaling pathway. PLoS One 2016; 11:e0164237.
- [172] Sun GY, Chen Z, Jasmer KJ, Chuang DY, Gu Z, Hannink M, Simonyi A. Quercetin attenuates
 inflammatory responses in BV-2 microglial cells: Role of MAPKs on the Nrf2 pathway and
 induction of heme oxygenase-1. PLoS One 2015; 10:e0141509.
- [173] Dai C, Li B, Zhou Y, Li D, Zhang S, Li H, Xiao X, Tang S. Curcumin attenuates quinocetone
 induced apoptosis and inflammation via the opposite modulation of Nrf2/HO-1 and NF-kB
 pathway in human hepatocyte L02 cells. Food Chem Toxicol 2016; 95:52-63.
- [174] Das L, Vinayak M. Curcumin modulates glycolytic metabolism and inflammatory cytokines via
 Nrf2 in dalton's lymphoma ascites cells in vivo. Anticancer Agents Med Chem 2018; doi:
 10.2174/1871520618666180604093802.
- [175] Han SG, Han SS, Toborek M, Hennig B. EGCG protects endothelial cells against PCB 126 induced inflammation through inhibition of AhR and induction of Nrf2-regulated genes. Toxicol
 Appl Pharmacol 2012; 261:181-8.
- [176] Lin CW, Wu MJ, Liu IY, Su JD, Yen JH. Neurotrophic and cytoprotective action of luteolin in
 PC12 cells through ERK-dependent induction of Nrf2-driven HO-1 expression. J Agric Food
 Chem 2010; 58:4477-86.
- 44 [177] Wang R, Tu J, Zhang Q, Zhang X, Zhu Y, Ma W, Cheng C, Brann DW, Yang F. Genistein
 45 attenuates ischemic oxidative damage and behavioral deficits via eNOS/Nrf2/HO-1 signaling.
 46 Hippocampus 2013; 23:634-47.
- 47 [178] Wang-Sheng C, Jie A, Jian-Jun L, Lan H, Zeng-Bao X, Chang-Qing L. Piperine attenuates
 48 lipopolysaccharide (LPS)-induced inflammatory responses in BV2 microglia. Int
 49 Immunopharmacol 2017; 42:44-8.

5 effects on oxidative status in animals - A review. Asian-Australas J Anim Sci 2017; 30:299-308. 6 Camacho-Barquero L, Villegas I, Sanchez-Calvo JM, Talero E, Sanchez-Fidalgo S, Motilva V, [181] 7 Alarcon de la Lastra C. Curcumin, a curcuma longa constituent, acts on MAPK p38 pathway 8 modulating COX-2 and iNOS expression in chronic experimental colitis. Int Immunopharmacol 9 2007; 7:333-42. 10 [182] Gao Y, Kang L, Li C, Wang X, Sun C, Li Q, Liu R, Wang J. Resveratrol ameliorates diabetesinduced cardiac dysfunction through AT1R-ERK/p38 MAPK signaling pathway. Cardiovasc 11 12 Toxicol 2016; 16:130-7. 13 Shen MY, Hsiao G, Liu CL, Fong TH, Lin KH, Chou DS, Sheu JR. Inhibitory mechanisms of [183] 14 resveratrol in platelet activation: pivotal roles of p38 MAPK and NO/cyclic GMP. Br J Haematol 15 2007: 139:475-85. 16 Nonn L, Duong D, Peehl DM. Chemopreventive anti-inflammatory activities of curcumin and [184] 17 other phytochemicals mediated by MAP kinase phosphatase-5 in prostate cells. Carcinogenesis 18 2007; 28:1188-96. 19 [185] Seo MJ, Lee YJ, Hwang JH, Kim KJ, Lee BY. The inhibitory effects of quercetin on obesity and 20 obesity-induced inflammation by regulation of MAPK signaling. J Nutr Biochem 2015; 26:1308-21 16 22 Yu L, Yu H, Li X, Jin C, Zhao Y, Xu S, Sheng X. P38 MAPK/miR-1 are involved in the [186] 23 protective effect of EGCG in high glucose-induced Cx43 downregulation in neonatal rat 24 cardiomyocytes. Cell Biol Int 2016; 40:934-42. 25 Subhashini, Chauhan PS, Dash D, Paul BN, Singh R. Intranasal curcumin ameliorates airway [187] 26 inflammation and obstruction by regulating MAPKinase activation (p38, Erk and JNK) and 27 prostaglandin D2 release in murine model of asthma. Int Immunopharmacol 2016; 31:200-6. 28 Eo H, Jeon YJ, Lee M, Lim Y. Brown Alga Ecklonia cava polyphenol extract ameliorates hepatic [188] 29 lipogenesis, oxidative stress, and inflammation by activation of AMPK and SIRT1 in high-fat 30 diet--induced obese mice. J Agric Food Chem 2015; 63:349-59. 31 Yang Z, Kahn BB, Shi H, Xue BZ. Macrophage alpha1 AMP-activated protein kinase [189] 32 (alpha1AMPK) antagonizes fatty acid-induced inflammation through SIRT1. J Biol Chem 2010; 33 285:19051-9. 34 Salminen A, Hyttinen JM, Kaarniranta K. AMP-activated protein kinase inhibits NF-kappaB [190] 35 signaling and inflammation: impact on healthspan and lifespan. J Mol Med (Berl) 2011; 89:667-36 76. 37 [191] Grisouard J, Dembinski K, Mayer D, Keller U, Muller B, Christ-Crain M. Targeting AMP-38 activated protein kinase in adipocytes to modulate obesity-related adipokine production 39 associated with insulin resistance and breast cancer cell proliferation. Diabetol Metab Syndr 2011; 40 3:16. 41 [192] Kaminska B. MAPK signalling pathways as molecular targets for anti-inflammatory therapy-42 from molecular mechanisms to therapeutic benefits. Biochim Biophys Acta 2005; 1754:253-62. 43 Zhang QY, Pan Y, Wang R, Kang LL, Xue QC, Wang XN, Kong LD. Quercetin inhibits [193] 44 AMPK/TXNIP activation and reduces inflammatory lesions to improve insulin signaling defect in 45 the hypothalamus of high fructose-fed rats. J Nutr Biochem 2014; 25:420-8. 46 [194] Kaneider NC, Mosheimer B, Reinisch N, Patsch JR, Wiedermann CJ. Inhibition of thrombin-

Yang M, Jiang ZH, Li CG, Zhu YJ, Li Z, Tang YZ, Ni CL. Apigenin prevents metabolic

syndrome in high-fructose diet-fed mice by Keap1-Nrf2 pathway. Biomed Pharmacother 2018;

Lee MT, Lin WC, Yu B, Lee TT. Antioxidant capacity of phytochemicals and their potential

1

2 3

4

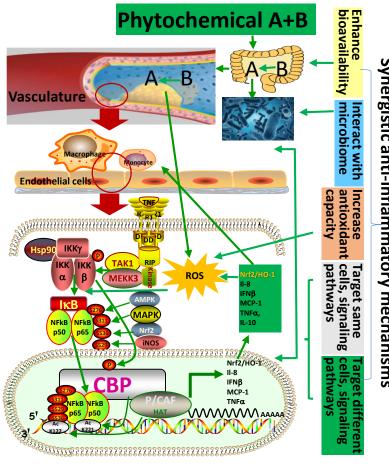
[179]

[180]

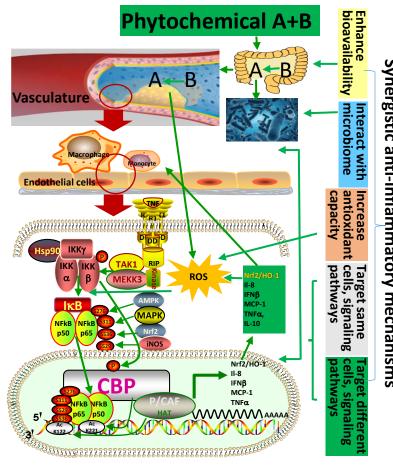
105:1283-90.

- induced signaling by resveratrol and quercetin: effects on adenosine nucleotide metabolism in endothelial cells and platelet-neutrophil interactions. Thromb Res 2004; 114:185-94.
 [195] Wang AY, Al-Kuhlani M, Johnston SC, Ojcius DM, Chou J, Dean D. Transcription factor
- (1)5] Wang AT, AFRaman M, Johnston SC, Ojetas DM, Chou J, Dean D. Hansenpuon factor
 complex AP-1 mediates inflammation initiated by Chlamydia pneumoniae infection. Cell
 Microbiol 2013; 15:779-94.

- [196] Andreadi CK, Howells LM, Atherfold PA, Manson MM. Involvement of Nrf2, p38, B-Raf, and nuclear factor-kappa B, but not phosphatidylinositol 3-kinase, in induction of hemeoxygenase-1 by dietary polyphenols. Mol Pharmacol 2006; 69:1033-40.



Synergistic anti-inflammatory mechanisms



Synergistic anti-inflammatory mechanisms